

Review

Mast cells in the autonomic nervous system and potential role in disorders with dysautonomia and neuroinflammation



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Key Messages

- Mast cells are located close to the blood vessels and neurons.
- Mast cells secrete many vasoactive and neurosensitizing mediators without histamine or tryptase.
- Mast cells can affect and be affected by the autonomic nervous system.
- Mast cells could contribute to dysautonomias and neuroinflammation.
- There is a need for development of effective inhibitors of mast cell activation.

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ABSTRACT

Mast cells (MC) are ubiquitous in the body, and they are critical for not only in allergic diseases but also in immunity and inflammation, including having potential involvement in the pathophysiology of dysautonomias and neuroinflammatory disorders. MC are located perivascularly close to nerve endings and sites such as the carotid bodies, heart, hypothalamus, the pineal gland, and the adrenal gland that would allow them not only to regulate but also to be affected by the autonomic nervous system (ANS). MC are stimulated not only by allergens but also many other triggers including some from the ANS that can affect MC release of neurosensitizing, proinflammatory, and vasoactive mediators. Hence, MC may be able to regulate homeostatic functions that seem to be dysfunctional in many conditions, such as postural orthostatic tachycardia syndrome, autism spectrum disorder, myalgic encephalomyelitis/chronic fatigue syndrome, and Long-COVID syndrome. The evidence indicates that there is a possible association between these conditions and diseases associated with MC activation. There is no effective treatment for any form of these conditions other than minimizing symptoms. Given the many ways MC could be activated and the numerous mediators released, it would be important to develop ways to inhibit stimulation of MC and the release of ANS-relevant mediators.

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Introduction

Mast cells (MC) are ubiquitous in the body,¹ and they are critical for allergic diseases.² MC disorders are divided in primary, including cutaneous and systemic mastocytosis, secondary, including atopic (IgE dependent such as asthma and atopic dermatitis) and other allergic diseases, and idiopathic, including MC activation disorders (MCADs).³ The latter were recently renamed “MCA-related disorders”

(still MCADs) and were subcategorized with “atopic diathesis and hypersensitivity disorders” listed under MCADs as “conditions predisposing to MCA/MCAD.”⁴ However, it is apparent that MCA can occur in many MC disorders in response to known or unknown triggers.⁵ To avoid any confusion, the term “diseases associated with mast cell activation” is used in this review presenting evidence linking MC and MCA with the autonomic nervous system (ANS) and discusses their possible involvement in the pathophysiology of certain conditions characterized by dysregulation of the ANS.

The role of MC in health and disease has been discussed extensively not only in allergies^{2,6–15} but also in immunity and

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inflammation.^{2,16–19} MC can secrete histamine and multiple proinflammatory cytokines and chemokines,^{15,20} including interleukin-1 beta (IL-1β),²¹ IL-6,²² IL-31,²³ and tumor necrosis factor (TNF).²⁴ Most reviews have focused on mastocytosis,^{3,25–29} but MC activation has been gaining interest because of its potential involvement in the pathophysiology of neuroinflammation⁵ and neuropsychiatric disorders,^{30,31} especially because patients with mastocytosis present with cognitive impairment^{32–34} that may be related to brain abnormalities.³⁵

MC respond not only to allergens but also to many other stimuli that can act alone or increase MC reactivity.³ When stimulated by IgE through aggregation of its high-affinity IgE surface receptor (FcεRI), MC can release several neurosensitizing, proinflammatory, and vasoactive mediators.^{36,37} Some of these mediators are prestored in secretory granules (eg, histamine, tryptase, and TNF)^{38,39} and are released immediately after stimulation. MC can also secrete mitochondrial DNA (mtDNA) extracellularly,⁴⁰ which serves as an alarmin and can stimulate proinflammatory mediator secretion from immune cells.^{41,42} Other mediators are newly synthesized such as chemokines (eg, C-C motif chemokine ligand 2 [CCL2], C-X-C motif chemokine ligand 8 [CXCL8/IL-8],⁴³ and cytokines [IL-6,²² IL-1β,²¹ and TNF]²⁴). Even though IL-6 is not specific for MC, it can be released selectively,^{22,44} was elevated in systemic mastocytosis, and correlated with disease severity.^{45–47} In fact, IL-6 promotes an increase in MC numbers.⁴⁸ MC can be activated by viruses.^{49,50} MC can also secrete anti-inflammatory molecules such as IL-10, transforming growth factor beta, and chondroitin sulfate.^{51–53}

In addition to allergic triggers, MC can be stimulated by nonallergic agents,^{2,54,55} especially neuropeptides,⁵⁶ such as substance P (SP)^{21,57} and the SP-related hemokinin-1 (HK-1),⁵⁸ which have proinflammatory properties.^{59–63} Moreover, SP induced the ST2 receptor for IL-33²⁴ further exacerbating MC activation. Under such conditions, especially when primed by the alarmin IL-3,^{55,64} MC can release mediators without histamine or tryptase,⁶⁵ a process by which MC can participate in inflammatory diseases.^{2,17}

The regulation of MC by neurotransmitters and neuropeptides has been reviewed,^{56,66,67} with emphasis on corticotropin-releasing hormone (CRH),³⁷ HK-1,⁵⁸ nerve growth factor,⁶⁸ neurotensin (NT),⁶⁹ SP,⁵⁷ and somatostatin^{70,71} acting by activation of high-affinity receptors. MC can also be stimulated by cationic molecules and drugs through activation of the low-affinity surface receptor Mas-related G-protein coupled receptor member X2 (MRGPRX2).^{72–74} Secretion of mediators can occur using different signaling^{14,75–77} and secretory^{75,77} pathways.

In addition to the connective tissue and mucosal location, MC are also located in many endocrine glands, including the hypothalamus, pineal, and pituitary glands (Table 1)⁵⁶; hence, they may be able to regulate homeostatic functions that seem to be dysregulated in many conditions, all of which worsen with psychological stress⁷⁸ and some of which are discussed subsequently.

However, the available evidence of the involvement of MC in the ANS is not uniform; it is often circumstantial, with many contradicting reports depending on the species and trigger. Moreover, human studies were based on a few subjects who are often not well defined.

Table 1
Anatomic Sites of Mast Cells Relevant to ANS

• Adrenal glands
• Blood-brain barrier
• Cardiac pacemakers
• Carotid bodies
• Coronary arteries
• Hypothalamus
• Pineal gland
• Vagus nerves
• Vasculature

Abbreviation: ANS, autonomic nervous system.

In particular, the definitions of MCAD and more so of MC activation syndrome (MCAS) are often not precise and are used interchangeably casting doubt on the validity of some associations reported, especially with dysautonomias.

Mast Cells in the Autonomic Nervous System

MC are located in close proximity to the blood vessels and peripheral nerves, which allow them to be strategically positioned to modulate sympathetic activity and vascular tone (Table 1). In fact, MC can communicate with neurons⁷⁹ by either direct contact,⁸⁰ through the nanotubes,⁸¹ or through release of relevant mediators. MC can also take up from the extracellular space and store and release additional mediators such as dopamine, epinephrine, norepinephrine, and serotonin (Table 2). MC could be affected by the ANS, especially at sites such as the carotid body,⁸² the coronary arteries,⁸³ and the heart (Table 1, Fig 1).^{84,85} MC could also affect the ANS in many different ways (Table 3) through activation of MC receptors by molecules released from the ANS (Fig 2) and through the release of ANS-acting MC mediators such as acetylcholine (ACh), angiotensins, calcitonin gene-related peptide, CRH, endorphins, HK-1, histamine, nerve growth factor, prostaglandin D2 (PGD₂), TNF-α, tryptase, and vascular endothelial growth factor (VEGF) (Table 2), to be discussed subsequently (Fig 2).

There is a long history of contradictory results concerning the effects of ACh, which could affect MC by both the muscarinic and nicotinic receptors,⁸⁶ but the published evidence is mostly based on rodent MC and on super pharmacologic concentrations of ACh or its agonists. In addition to the vagus nerve,⁸⁷ there are many non-neuronal sources of ACh such as epithelial, endothelial, and immune cells, including MC, themselves.⁸⁸ In humans, Ch was increased as much as 14-fold in the epidermis of skin biopsies from patients with atopic dermatitis (AD), implying it must be somehow involved.⁸⁹ One article reported that ACh stimulated the skin only in the cholinergic urticaria type with anhidrosis or hyperhidrosis through activation of M3 cholinergic receptors.⁹⁰

It was originally reported that ACh could stimulate histamine release from abdominal and thoracic MC of Wistar albino rats at concentrations as low as 10^{−10} M.⁹¹ However, subsequent publications using different types of rodents did not confirm those original findings. For instance, the ACh agonist pilocarpine stimulated histamine release from rat peritoneal MC, but only at a concentration of 10^{−2} M,⁹² whereas another article reported that ACh could not stimulate histamine release from rat MC even at 10^{−2} M.⁹³ The ACh analogue carbachol (10^{−4} M) did not stimulate rat peritoneal MC histamine release but augmented the effect of SP.⁹⁴ Instead, other scientists reported that carbachol neither stimulated nor augmented histamine release from rat peritoneal MC.⁹⁵

Table 2
ANS-Relevant Mast Cell Mediators^a

• ACh
• Angiotensins
• CGRP
• CRH
• Endorphins
• HK-1
• Histamine
• NGF
• PGD ₂
• TNF-α
• Tryptase
• VEGF

Abbreviations: ANS, autonomic nervous system; ACh, acetylcholine; CGRP, calcitonin gene-related peptide; CRH, corticotropin-releasing hormone; NGF, nerve growth factor; PGD₂, prostaglandin D2; TNF-α, tumor necrosis factor-alpha; VEGF, vascular endothelial growth factor.

^aMast cells can also internalize from the extracellular space and store and release additional mediators such as dopamine, epinephrine, norepinephrine, and serotonin.

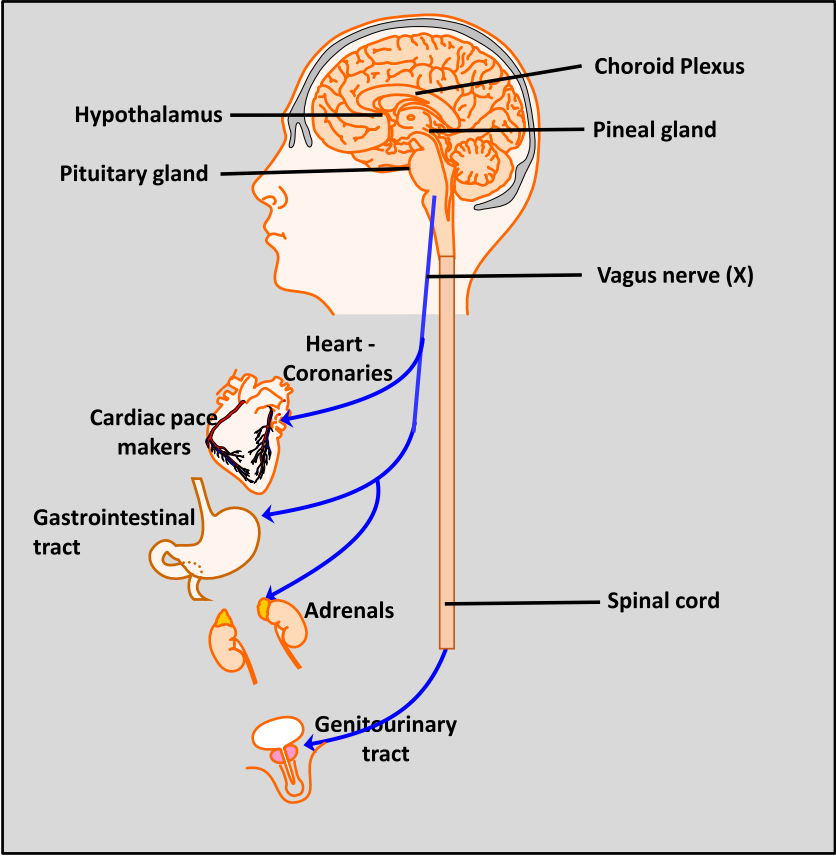


Figure 1. Diagrammatic representation of anatomic sites where mast cells may interact with the ANS. ANS, autonomic nervous system.

In contrast to the Ach effect through activation of the muscarinic receptors, activation of the nicotinic receptors may have the opposite effect. For instance, Ach inhibited histamine release from isolated human bronchi.⁹⁶ Mucosal MC activation was also found to be negatively regulated through the $\alpha 7$ nicotinic receptor subunit.⁹⁷ The $\alpha 7$ nicotinic receptor subunit, which has anti-inflammatory effects, was significantly elevated in a mouse model of stress.⁹⁸ Activation of nicotinic receptors inhibited vagus-stimulated histamine release in a murine food allergy model,⁹⁹ including the allergy-induced release of leukotrienes and cytokines, but not release of histamine or degranulation.¹⁰⁰ The same was true in mucosal-like MC.⁹⁷ Adrenergic agents could also inhibit Ach-stimulated histamine release from isolated rat MC through activation of the beta-adrenoreceptor.¹⁰¹ However, the clinical effects of adrenergic drugs are more complex mostly due to their actions on other organs/tissues. Adrenergic agonists such as terbutaline dilate the bronchi, and intramuscular epinephrine is used to counter bronchoconstriction and hypertension associated with anaphylaxis. In contrast, the beta-blocker propranolol is useful for anxiety, palpitations, and migraines often experienced by patients with

MCADs. Hence, the effect of the parasympathetic system may depend on the relative MC expression of muscarinic and nicotinic receptors, including adrenoreceptors, which may vary depending on species, anatomic location, and disease activity.

There is additional evidence that MC may participate in autonomic function and dysfunction. MC are located close to and communicate with the pineal gland¹⁰² and may both regulate and be affected by the circadian clock.^{103–105} Moreover, extrapineal melatonin was identified in platelets and endothelial and immune cells, including MC.¹⁰⁶ A thermoregulatory neuron circuit axis was reported through MC-derived chymase leading to hypothermia during anaphylaxis.¹⁰⁷ Post-exercise hypotension and collapse were linked to MC.¹⁰⁸ Decreased cerebral blood flow and small fiber

Table 3
ANS-Associated Mast Cell Actions

- Activation of microglia and astrocytes
- BBB disruption
- Cardiac pacemaking
- Circadian clock
- Coronary artery constriction
- HPA axis regulation
- Neuroinflammation
- Neurosensitization
- Vascular tone control

Abbreviations: ANS, autonomic nervous system; BBB, blood-brain barrier; HPA, hypothalamus-pituitary-adrenal.

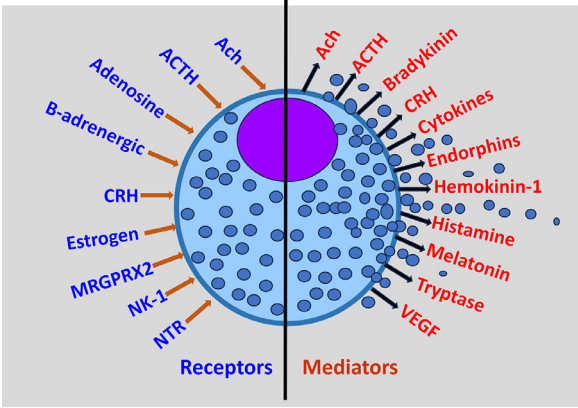


Figure 2. Diagrammatic representation of receptors and mediators relevant to the ANS. Ach, acetylcholine; ACTH, adrenocorticotropic hormone; ANS, autonomic nervous system; CRH, corticotropin-releasing hormone; VEGF, vascular endothelial growth factor.

neuropathy were found to be present in patients with systemic mastocytosis.¹⁰⁹

Mast Cells in the Central Nervous System

MC are present in the CNS perivascularly,^{110,111} especially in the meninges,^{80,112} and the median eminence¹¹³ of the hypothalamus.^{80,114,115} The presence of an increased number of MC-containing histamine has been reported in the human brain,¹¹⁶ especially in brain lesions of patients with multiple sclerosis (MS).^{117–120} We had called brain MC the “immune gate to the brain.”¹¹⁰ We reported increased levels of MC tryptase in the cerebrospinal fluid of patients with MS.¹²¹ Interestingly, a letter to the editor reported that the tryptase level in the cerebrospinal fluid of 4 patients with mastocytosis was not elevated,¹²² but none of these patients apparently had neurologic symptoms and the serum tryptase level of one of the 4 patients was not even elevated (3.2 ng/mL).

Stimulation of the brain and nasal MC resulted in stimulation of the hypothalamus-pituitary-adrenal (HPA) axis,^{37,123–126} possibly through MC release of histamine,¹²⁷ IL-6,^{127,128} and CRH,¹²⁹ implying that either these mediators from the nasal MC reached the hypothalamus or the antigen reached the hypothalamic MC. In addition, perivascular brain MC could be stimulated by various triggers including environmental and food allergens leading to disruption of the blood-brain barrier (BBB) and subsequent entry of additional triggers in the brain. In fact, MC-derived histamine,^{130–133} tryptase,¹³⁴ and cytokines^{135,136} increased BBB permeability,^{114,133,137} thus permitting MC mediators and other molecules to enter the brain parenchyma.^{138,139} In particular, prostaglandin E2 was reported to cross the BBB.¹⁴⁰

Functional interactions have been reported between MC and neurons^{80,141,142} that are often positive for CRH^{80,143} secreted under stress.⁸⁰ The effect of stress on MC has also been reviewed.^{78,144} Restraint stress in rodents increased BBB permeability^{114,137,145} through CRH-stimulating MC.^{137,146,147} Restraint stress also led to increased mammary adenocarcinoma brain metastases in mice.¹⁴⁶ This process could worsen with stress acting through CRH stimulation of MC,^{137,146} leading to increased dura vascular permeability. We found that stress stimulates MC^{78,144} through CRH⁷⁸ leading to increased dura vascular permeability, an effect that was absent in MC-deficient mice.¹⁴⁸ Interestingly, meningeal MC affected the integrity of the BBB and promoted T-cell brain infiltration.¹⁴⁹ MC were found to be an early activator of lipopolysaccharide (LPS)-induced neuroinflammation and BBB dysfunction in the hippocampus.¹⁵⁰ Stress has also been linked to the possible priming of immune cells, thus contributing to neuroinflammation in Alzheimer’s disease.^{151,152}

Moreover, NT¹⁵³ and SP,¹⁵⁴ neuropeptides implicated in inflammation, induced CRHR-1, thus creating an autocrine loop. Furthermore, SP induced the ST2 receptor for IL-33²⁴ further exacerbating MC activation by the combined action of neuropeptides and IL-33.

Restraint stress in rodents increased BBB permeability^{114,137,145} through CRH-stimulating MC.^{137,146,147} MC-derived mediators, such as cytokines,^{135,136} increased BBB permeability not only to small molecules^{114,137} but also to mammary adenocarcinoma brain metastases in mice.¹⁴⁶ This process could worsen with stress acting through CRH activation of MC,^{137,146} leading to increased dura vascular permeability. Interestingly, meningeal MC affected the integrity of the BBB and promoted T-cell brain infiltration.¹⁴⁹ MC were found to be an early activator of LPS-induced neuroinflammation and BBB dysfunction in the hippocampus.¹⁵⁰ MC responsiveness may be regulated by the neuroimmune stimuli and effects of the different receptors involved. For instance, MC expresses high-affinity neurokinin-1 receptors for SP.^{24,154,155} Moreover, SP¹⁵⁴ and NT¹⁵³ induced the expression of CRHR-1 in human MC. MC are the richest source of histamine in the amygdala, hippocampus, hypothalamus, and thalamus.¹⁵⁶ Neuronal histamine is involved in neurodevelopment¹⁵⁷ and acts as an alert signal in the brain.¹⁵⁸ In fact, brain histamine is critical for arousal,¹⁵⁹ memory consolidation and retrieval,¹⁶⁰ and motivation.^{156,159–167}

However, extensive release of histamine from brain MC or peripheral histamine crossing a disrupted BBB could have opposite results. Activated brain MC were found to contribute to postoperative cognitive dysfunction by activating microglia¹⁶⁸ through the release of mediators.¹⁶⁹ Activation of MC^{151,170} and microglia¹⁷¹ in the hypothalamus¹⁷² could lead to dysfunctional homeostasis and cognitive dysfunction,¹⁶⁸ most often also found in patients with MCADs,^{34,173} and may be related to brain abnormalities.

Mast Cells Communicate With Microglia in Neuroinflammation

Microglia have been considered key players in brain injury¹⁷⁴ and the development of complex^{175,176} neuroinflammatory^{175,176} and neurodegenerative^{177–184} disorders. Microglia express toll-like receptors¹⁸⁵ activated by damage-associated molecular patterns and pathogen-associated molecular patterns. Microglia are typically characterized as proinflammatory (M1) and anti-inflammatory (M2).¹⁸⁶ On stimulation, activated microglia adopt an amoeboid shape (M1) that renders them capable of phagocytosis and antigen presentation, including the release of proinflammatory cytokines, free radicals, and fatty acid metabolites.¹⁸⁷ Microglia also communicate with astrocytes.¹⁸⁸ As a result, microglia have been found to play a key role in neuroinflammation and neurodegenerative diseases.^{115,189} An important consequence of microglial priming is that the cytokines and chemokines released can induce astrogliosis, amyloid deposition, and subsequently, further worsening neuroinflammation.^{190–192} Microglia express receptors for NT,¹⁹³ and we reported that NT can activate human microglia to secrete proinflammatory molecules.¹⁹⁴ Microglia also express receptors for CRH, secreted under stress.¹⁹⁵ Psychological stress can increase microglial reactivity to other challenges¹⁹⁶ and lead to cognitive decline.¹⁹⁷

MC communicate with microglia^{198,199} leading to their activation^{169,198,200,201} and contributing to neuroinflammation^{199–202} and neurodegenerative diseases.^{199,203,204} This effect is absent in MC-deficient mice.^{203,205,206} Microglia can be activated by molecules released from MC, such as histamine²⁰⁷ and tryptase.²⁰⁸ In fact, MC proteases can activate astrocytes and glia/neurons to release IL-33.²⁰⁹ Stabilization of MC was found to inhibit LPS-induced neuroinflammation by inhibiting activation of the microglia.²¹⁰ Food allergy that depends on MC activation has also been found to increase both the total and the activated microglia, including levels of TNF, in the hippocampus associated with behavioral and learning impairments.¹⁶⁷

Mast Cells in Ehlers-Danlos Syndrome/Postural Orthostatic Tachycardia Syndrome

MC activation-associated disorders have also been reported in conditions characterized by dysfunction of the ANS, such as cholinergic urticaria and possibly postural orthostatic tachycardia syndrome (POTS),^{211,212} patients who experience flushing in addition to tachycardia, and Ehlers-Danlos syndrome (EDS).^{213,214} In such cases, MC activation may be compensated by hyperadrenergic stimulation.²¹⁵ Sporadic reports suggested that some of these patients may even present with the “triad” of MCAS, POTS, and EDS.²¹⁶ One retrospective study reviewed records of patients with EDS and POTS (n = 195) and reported that 31% of these patients had MCAS as compared with 2% for the healthy control (Table 4)²¹²; however, the criteria for defining MCAS were not clear.

One retrospective case series (n = 98) reported that 24% of patients with EDS had MCA.²¹⁷ Other papers reported on the comorbidity of other MC activation-associated diseases. One retrospective case series of patients with either hypermobility syndrome (n = 126) or EDS (n = 162) as compared with healthy controls (n = 221) revealed that there was a significant increase in the frequency of respiratory symptoms in subjects with both hypermobility syndrome

Table 4
Characteristics of Studies Reviewed

Type of study	Comorbidity ^a	Diagnosis	Evaluation	No patients	Findings	Conclusions
EDS, POTS						
Prospective	MCA	Hyperadrenergic POTS	Clinical examination	177		MCA was common ²¹¹
Prospective	Asthma	Joint hypermobility	Clinical examination	Asthma (n = 100) Healthy controls (n = 100)	Hypermobility was present in 70.1% of patients with asthma as compared with 29.9% in controls	Hypermobility was common in patients with asthma ²¹⁶
Prospective	Asthma, atopic conditions	HMS, EDS	Questionnaires	509 subjects (126 HMS, 162 EDS, and 221 healthy controls)	HMS(asthma OR, 2.4; 95% CI, 1.4–4.1, $P = .002$ and atopy OR, 2.7; 95% CI, 1.6–4.5, $P < .001$) and EDS (asthma OR, 3.1; 95% CI, 1.8–5.2, $P < .001$ and atopy OR, 2.6; 95% CI, 1.6–4.4, $P < .001$) compared with controls	Asthma and atopic conditions were more frequent in patients with HMS and EDS ²¹⁵
Retrospective	MCAS	POTS + EGID	EMRs	7	Autonomic dysfunction was 5- to 10-fold higher in EGID than expected in the general population	2 Subjects (29%) had comorbid EDS and MCAS ²¹⁷
Retrospective	MCAS	EDS + POTS	Record review—diagnostic codes	195	31% vs 2% controls	MCAS was more common ²⁰⁸
Retrospective	MCA	MCA + POTS	Record review—diagnostic codes	69	42% of patients with POTS had elevated biochemical markers suggesting MCA disorder.	MCA disorder is common in POTS ²¹⁸
Prospective	MC	Connective tissue disorders	Skin biopsies	12 Patients and 16 controls	1.7-fold greater number of chymase-positive skin MC	MC may be related to arterial hypertension in the patients ²¹⁹
ASD						
Retrospective	Allergies,mastocytosis, UP	ASD	Questionnaire	41	10 × higher	MCAD increased the risk for ASD ²²⁰
Retrospective	Asthma	ASD	2007 National Survey of Children's Health Data Base	77,951	OR, 1.19; 95% CI, 1.03–1.36	Asthma was 35% more common in ASD ²²¹
Retrospective	Food allergies	ASD	CHARGE study	ASD (n = 560), control (n = 391)	OR, 2.23; 95% CI, 1.28–3.89	Food allergies are more common in ASD ²²²
Systematic review	AD	ASD	18 Studies up to 2015		HR, 3.4; OR, 1.24 ($P < .001$)	AD increased the risk for ASD ²²³
Retrospective	AD	ASD	Taiwan's National Health Insurance Program (2000–2010)	387,262 with AD	HR: 1.75	AD before 2 y old increased the risk of AD ²²⁴
Retrospective	Food allergy	ASD	Population-based, cross-sectional study using data from the National Health Interview Survey (1997–2016)	199,520	OR, 2.29; 95% CI, 1.87–2.81	Food allergy increased the risk for ASD ²²⁵
Retrospective	Food allergies	ASD	National Health Interview Survey (2011–2015)	53,365 of whom 905 with ASD and 2977 with food allergies	2.5 higher ASD in the presence of food allergies	Food allergies increased the risk for ASD ²²⁶
Retrospective	Food hypersensitivities	ASD	8 Data bases before 2020	434,809	OR, 2.79; OR, 2083.75 ($P < 0.1$)	Food sensitivities increased the risk for ASD ²²⁷
Prospective	AD	ASD	Record review	446	OR, 4.26; 95% CI, 1.36–13.36	AD increased the risk for ASD ²²⁸
Prospective	Asthma, allergies, eczema, and hay fever	ASD	Clinical assessment	45 with ASD vs 93 controls	2.4 times higher	Atopy increases the risk for ASD ²²⁹
Retrospective	AD	ASD	UK health records data	409,431 with AD vs 1,809,029 without AD	HR, 1.02; 95% CI, 0.98–1.06	AD increases the risk for ASD ²³⁰
Retrospective	Food allergy	ASD	Meta-analysis included 12 published articles with 434,809 subjects.	434,809	OR, 2.79; 95% CI, 2.01–3.75	Food allergy increases the risk for ASD ²²⁷
Retrospective	Allergic disorders	ASD	Pediatric database of Clalit Health Services (2000–2018)	117,022 with allergies vs 116,968 without allergies	OR, 1.17; 95% CI, 1.08–1.27	Allergic disorders increase the risk for ASD ²³¹
Retrospective	Asthma	ASD	Familial co-aggregation of asthma and ASD in individuals born in Sweden (1992–2007)	1,569,944 including their full- and half-siblings (1,704,388 and 356,544 pairs)	OR, 1.33; 95% CI, 1.28–1.37 and OR, 1.44; 95% CI, 1.38–1.50, for full siblings	Asthma increased the risk for ASD ²³²

(continued)

Table 4 (Continued)

Type of study	Comorbidity ^a	Diagnosis	Evaluation	No patients	Findings	Conclusions
Prospective	Asthma and other atopic conditions	ASD	Enrolled at birth and followed clinically	2580 Children enrolled at birth were followed prospectively, of which 119 have ASD vs 1273 controls	OR, 1.38; 95% CI, 1.15-1.66 for asthma and OR, 1.52; 95% CI, 1.28-1.81 for combined atopic conditions	Asthma increased the risk for ASD that increased further in the presence of multiple atopic conditions ²³³
ME/CFS Prospective	Allergies	CFS	Clinical assessment and blood cytokines	CFS (n = 18) Allergic (n = 14) Control (n = 11)	15/18 Patients with CFS have allergy	Allergies were more common in patients with CFS ²³⁴
Retrospective	MCA	ME/CFS	Patients with CFS/ME were defined in accordance with the International Consensus Criteria; blood samples collected	31	There was a significant increase in CD40 ligand and MHC-II receptors on differentiated MC populations in severe CFS/ME compared with healthy controls and moderate CFS/ME.	Significant increase of naive MC in patients with moderate and severe CFS/ME compared with healthy controls ²³⁵
Retrospective		EDS	Rehabilitation clinic	98	24% of patients with EDS had MCAS	MCAS was more common in patients with EDS ²¹⁴
COVID-19 Retrospective	MC	COVID-19	Lung autopsies	COVID = 6 and HC = 10	Increased perivascular and septal CD117 + MC	Increased perivascular and septal MC in the lungs ²³⁶
Retrospective	MC-associated proteases	COVID-19	Lung autopsies and serum	COVID = 19 and HC = 20	Proteases were significantly increased in the serum and their gene expression in the lungs	Increased MC-associated proteases in the serum and lungs of patients with COVID-19 ²³⁷
Retrospective	Chymase	COVID-19	Plasma	Intubated = 7 Inpatient = 18 Community = 12 HC = 13	Chymase levels were increased in hospitalized patients compared with mild community patients	Increased serum chymase correlated with disease severity ²³⁸
Retrospective	MC	COVID-19	Lung autopsies with or without DAD	COVID = 19 Controls = 4	Organizing DAD in COVID-19 lungs revealed a 3-fold increase in MC between low- and high-density areas	Chymase and tryptase-positive MC were increased only in lung areas with DAD ²³⁹
Retrospective	MC were identified by KIT (CD117), tryptase, and chymase immunohistochemistry	COVID-19	Lung autopsies	30	Increased numbers of chymase-positive MC at early stage and further augmentation of MC during late stage of alveolar damage compared with controls	Increased number of MC in the lungs of patients with COVID-19 ²⁴⁰
Retrospective	MC-associated chymase, tryptase and carboxypeptidase A3	COVID-19	Lung autopsies and serum	COVID = 60 and HC = 17	All 3 proteases were increased in the serum and the density of degranulated MC was increased compared with control lungs.	SARS-CoV-2 was strongly associated with activation of MC ²⁴¹
Long-COVID Retrospective	MCA	Long-COVID	Online assessment, MCMRS score	Long-COVID = 136 MCA = 80 HC = 136	Long-COVID = 136 MCAS = 80 MCMRS score in patients with Long-COVID resembled that of patients with MCAS before treatment	MCA symptoms were increased in Long-COVID and mimicked the symptoms and severity reported by patients who had MCAS ²⁴²
Retrospective	Cytokines and proteases	PASC	Serum	PASC = 13 PAAC = 13 HC = 20	Chymase, tryptase, and carboxypeptidase A3 levels were elevated in active COVID-19, but only tryptase level remained elevated in PASC	MC-associated proteases are differentially expressed in active COVID-19 as compared with PASC ²⁴³

Abbreviations: AD, atopic dermatitis; ASD, autism spectrum disorder; CHARGE, Childhood Autism Risks from Genetics and Environment; CI, confidence interval; DAD, diffuse alveolar damage; EDS, Ehlers-Danlos syndrome; EGID, eosinophilic gastrointestinal disease; EMR, electronic medical record; HC, healthy control; HMS, hypermobility syndrome; ME/CFS, myalgic encephalomyelitis/chronic fatigue syndrome; MC, mast cell; MCA, mast cell activation; MCAS, mast cell activation syndrome; MCMRS, mast cell mediator release syndrome score; MC mediator release syndrome; OR, odds ratio; PAAC, post-acute asymptomatic COVID; PASC, post-acute symptomatic COVID; POTS, postural orthostatic tachycardia syndrome.

^aThe comorbidities are listed as indicated by the authors of the respective publications.

(asthma odds ratio [OR], 2.4; 95% CI, 1.4–4.1; $P = .002$ and atopy OR, 2.7; 95% CI, 1.6–4.5; $P < .001$) and EDS (asthma OR, 3.1; 95% CI, 1.8–5.2; $P < .001$ and atopy OR, 2.6; 95% CI, 1.6–4.4; $P < .001$) compared with controls (Table 4).²¹⁸ In another study, joint hypermobility was present in 70.1% of patients with asthma ($n = 100$) as compared with 29.9% in healthy controls ($n = 100$).²⁴⁴ In a small retrospective study of 7 patients with POTS and eosinophilic gastrointestinal disease, which is associated with activated MC, autonomic dysfunction was 5- to 10-fold higher in eosinophilic gastrointestinal disease than expected in the general population; moreover, 2 subjects (29%) had comorbid EDS and MCAS.²¹⁹ Another retrospective study of 69 patients with POTS found that 42% had elevated levels of biochemical markers suggesting the presence of MCA.²⁴⁵

One review²⁴⁶ summarized several of these papers and included 2 abstracts, one by Szalewski²⁴⁷ (2019) reporting an increased prevalence of idiopathic urticaria in patients with EDS and another by Cheung and Vadas²¹⁶ (2015) reporting that in patients with EDS ($n = 198$) from a rehabilitation center, 24% had a diagnosis of MCAS, but the criteria for this diagnosis were not clear.

Unfortunately, the definitions of MCAD and especially of MCAS were often not precise and used interchangeably, and some studies made inferences from the symptoms described in medical records casting doubt on the validity of some associations reported.²⁴⁸

Mast Cells in Autism Spectrum Disorder

Autism spectrum disorder (ASD) is characterized by difficulties in communication and apparently purposeless repetitive movements, cognitive impairment,²⁴⁹ but also extreme sensory sensitivities.²⁵⁰ The prevalence of ASD is estimated to be approximately 5% in the United States.²⁵¹ However, ASD pathogenesis is still unknown. In addition, most children with ASD have a number of comorbidities such as hyperactivity, gastrointestinal problems, allergies, and seizures,²⁵² making objective diagnosis and the development of effective treatments difficult.

We advanced the premise that focal inflammation in the amygdala due to MC and microglia activation can reset the fear threshold and throw the body into a perpetual fight-or-flight mode, thus “neglecting” higher functions.^{206,220,253,254} Using a questionnaire, we had reported that children born to mothers with systemic mastocytosis or with cutaneous mastocytosis themselves ($n = 40$) had an almost 10-fold higher risk of developing ASD²⁵⁵ (Table 4). Parental history of AD was also strongly associated with children developing AD.²⁵⁶ It had been reported that maternal immune activation²⁵⁷ and autoimmune diseases,²⁵⁸ especially psoriasis, but also allergies and asthma, were associated with a higher risk of ASD.²⁵⁹ In another study, almost 50% of children with ASD reported having relatives with rheumatoid diseases as compared with 26% in the control group.²⁶⁰ In a large study, mothers who experienced asthma, allergy, atopy, or eczema during pregnancy was associated with a higher risk of neuropsychiatric problems in the offspring.²⁶¹ A meta-analysis of 119 studies revealed that patients with parental history of atopic diseases had an increased risk for ASD (OR, 1.81; 95% CI, 1.65–1.99, $P < .001$).²⁵⁶

A recent publication revealed that the mother's circulating immune IgE resulted in vertical transmission of AD in the newborn through stimulation of fetal MC²⁶²; both passive and active prenatal sensitization conferred allergen sensitivity.²⁶² This important study indicated that fetal MC were functional and could be stimulated by specific IgE and allergens present in the mother during gestation.²⁶² Fetal MC could potentially respond to other stimuli such as neuropeptides and toxins, including the alarmin IL-33,^{55,263} with detrimental effects on brain development especially in premature babies.²⁶⁴ Inflammation during gestation could adversely affect the developing brain,^{265,266} including cortical dendritic arborization.²⁶⁷ In fact, prenatal allergen exposure was even found to program lifelong changes

in the social and sexual behaviors of adult rats, including effects on microglia activation and neonatal dendritic spine density.²⁶⁸

Subsequently, several large epidemiologic studies reported a strong association between AD and behavioral problems in general²⁶⁹ and ASD in particular.^{224,270–275} (Table 4). A strong association was reported between risk for developing ASD and allergies,^{220–223,274} especially asthma^{223,228} and AD,²³⁰ which could lead to brain inflammation and cognitive impairment.¹⁶⁷ One retrospective study reviewed 77,951 patients from the National Survey of Children's Health Data Set and found that asthma was 35% more common in ASD (OR, 1.19; 95% CI, 1.03–1.36).²²⁸ Another retrospective study investigated the effect of maternally reported asthma and allergies from the Childhood Autism Risks Genetics and Environment (CHARGE) study of 560 patients as compared with 391 healthy controls and found that food allergies were more common in ASD (OR, 2.23; 95% CI, 1.28–3.89) and correlated with the severity of symptoms.²²³ Another retrospective study using Taiwan's National Health Insurance Program (2000–2010) reviewed 387,262 children with AD and found that there was an increased risk for ASD (hazard ratio [HR], 1.75).²⁷⁴ A systematic review of 18 studies also found that AD increased the risk for ASD (HR, 3.4; OR, 1.24; $P < .001$).²³⁰

Additional studies confirmed the increased risk of ASD by the presence of atopic conditions (Table 4). One study reviewed records from 444 subjects and concluded that AD increased the risk of ASD (OR, 4.26; 95% CI, 1.36–13.36, $P < .001$).²²⁹ Another study investigated 409,431 patients with AD as compared with 1,809,029 subjects without AD and found that the presence of AD increased the risk of ASD (HR, 1.02; 95% CI, 0.98–1.06, $P < .001$).²³¹ (Table 4). One more retrospective study using the pediatric database of Clalit Health Services (2000–2018) evaluated 117,022 children with allergies as compared with 116,968 without allergies and concluded that allergic disorders increased the risk for ASD (OR, 1.17; 95% CI, 1.08–1.27, $P < .001$).²³² (Table 4). Another retrospective study investigated familial co-aggregation of asthma and ASD in individuals born in Sweden (1992–2007); the results from 1,569,944 children including their full- and half-siblings (1,704,388 and 356,544 pairs) were that asthma increased the risk for ASD which increased further in the presence of multiple atopic conditions (OR, 1.38; 95% CI, 1.15–1.66, for asthma, and OR, 1.52; 95% CI, 1.28–1.81 for combined atopic conditions, $P < .001$).²³³ (Table 4). A prospective study of asthma and other atopic conditions that investigated 2580 children enrolled at birth and followed clinically, of which 119 had ASD compared with 1273 healthy controls, concluded that asthma increased the risk for ASD which increased further in the presence of multiple atopic conditions (OR, 1.38; 95% CI, 1.15–1.66, for asthma, and OR, 1.52; 95% CI, 1.28–1.81 for combined atopic conditions)²⁷⁶ (Table 4). One more prospective study of 45 patients with ASD and 93 healthy controls concluded that the presence of asthma, allergies, eczema, or hay fever increased 2.4 times the risk of ASD²⁷⁷ (Table 4).

Food allergies and food intolerance also apparently increased the risk of ASD^{225–227,278–280} (Table 4). A population-based, cross-sectional study using data from the National Health Interview Survey (1997–2016) evaluated 199,520 subjects and concluded that food allergies increased the risk of ASD (OR, 2.29; 95% CI, 1.87–2.81; $P < .001$).²⁷⁸ (Table 4). In a retrospective study of parent-reported food allergies using the National Health Interview Survey (2011–2015) of 53,315 children, there were 905 patients with ASD and 2977 patients with food allergies resulting in 25 times more common food allergies in ASD (Table 4).²⁷⁹ One meta-analysis of 12 studies with a total of 434,809 subjects concluded that the presence of food hypersensitivities increased the risk of ASD (OR, 2.79; 95% CI, 2.08–3.75; $P < .001$).²²⁶ (Table 4). In fact, the presence of allergies was associated with elevated serum levels of autoantibodies against brain antigens in children with ASD.²⁸¹ It should be noted that parent-reported food allergies and sensitivities may also include food intolerances such as for histamine and phenols that are associated with dysfunctional

catabolic enzymes, such as diamine oxidase and catecholamine-ortho-methyl transferase, respectively.

Clearly, these results do not provide evidence of causation but certainly suggest some involvement of MCA at least in atopic diseases and ASD.

Mast Cells in Myalgic Encephalomyelitis/Chronic Fatigue Syndrome

Myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS) is a complex chronic, debilitating disorder^{282,283} with 4:1 female-to-male ratio,²³⁵ characterized by unrelenting fatigue, and autonomic, cognitive, and sleep dysfunction.^{172,284–286} especially physical and mental fatigue.²⁸⁷ The prevalence of ME/CFS is approximately 1% in the United States.^{235,288–291} Approximately half of the patients with ME/CFS develop symptoms after a sudden, influenza-like illness,²⁹² implying the possible involvement of pathogen-associated molecular patterns, including mycotoxins, such as ochratoxin A (OTA).²⁹³ As a result, ME/CFS may have an autoimmune component to its pathophysiology,^{294,295} involving neuroinflammation.^{296–298} Even though serum levels of proinflammatory cytokines, especially IL-1 β and IL-6,^{299–304} have been reported to be increased in patients with ME/CFS,^{304,305} other studies have not supported such findings,^{289,306} except maybe for elevated IL-6 level.^{304,307} In addition, there was no difference in cytokine content of extracellular vesicles from patient with ME/CFS as compared with healthy controls even though the number of vesicles was increased.³⁰⁸

Although the exact cause of ME/CFS remains unclear,³⁰⁹ evidence points to the involvement of some pathogenetic mechanism(s),^{303,310–312} such as neurovascular inflammation,^{112,200,313} endothelial dysfunction, BBB disruption, and reduced blood flow to the brain.^{314–318} Anti-receptor antibodies³¹⁹ and autoimmune gene expression³²⁰ have also been reported because MC-derived inflammatory, vasoactive, and neurotoxic mediators can disrupt the BBB,^{110,134} alter blood flow to the brain,¹⁰⁹ and can activate microglia,^{199,200} leading to neurovascular inflammation.¹⁵¹ MC have also been linked to fatigue in patients with mastocytosis³²¹ and neuropsychiatric symptoms.^{33,322} One study reported increased MC precursors in the blood of patients with ME/CFS,²⁸⁴ and another study revealed that the higher presence of atopic symptoms increases the risk of ME/CFS.^{323–326} A small prospective study investigated patients with MC/CFS and found that allergies were present in 15 of 18 patients (Table 4), which is higher than the general population (less than 30%).²³⁴ In this study, allergy was defined as “the presence of a wheal of 3 mm or greater in diameter on skin prick testing to one or more antigens in the standard Colorado inhalant panel and the presence of an atopic disease (allergic rhinitis, asthma, or atopic dermatitis).”²³⁴

We further revealed that cell-free mtDNA was increased in extracellular vesicles obtained from the blood of patients with ME/CFS³²⁷ and can amplify neuroinflammation.³²⁸ We reported that cultured human microglia can be stimulated by NT to release IL-1 β and CXCL8/IL-8.¹⁹⁴ This action was mediated through the regulatory complex mammalian target of rapamycin (mTOR),^{194,329} which has been implicated in brain development,³³⁰ aging,³³¹ dementia,³³² neuropsychiatric disorders,^{333–339} brain injury,³⁴⁰ neuroinflammation,³⁴¹ and central fatigue.³⁴²

Mast Cells in COVID-19 and Long-COVID Syndrome

Infection with SARS-CoV-2 leads to COVID-19, the severity of which derives from the host's immune response,³⁴³ especially the release of a storm of proinflammatory cytokines,^{344–351} such as IL-6,^{352–356} known as cytokine storms.³⁵⁷ Moreover, circulating mtDNA which is secreted from activated MC⁴⁰ was reported to be an indicator of severe illness and mortality in patients with COVID-19, as was extracellular mtDNA.³⁵⁸

A number of human and animal studies discussed MCA associated with SARS-CoV-2 infection or in COVID-19.^{236–238,241–243,359–368} In particular, one study of a few patients with COVID-19 (n = 6) reported increased MC density of CD117+ MC in the lungs of autopsies from patients with COVID-19 as compared with deceased controls (n = 10)²⁴³ (Table 4). Another article reported an increased number of chymase-positive MC in lung biopsy samples of patients with COVID-19 (n = 30)³⁶⁹ (Table 4).

Two papers reported increased serum levels of chymase in patients with COVID-19.^{237,241,368} In the second study, serum levels of chymase correlated with disease severity²³⁷ (Table 4). A third article reported that levels of chymase, tryptase, and carboxypeptidase A3 were all elevated in the serum of patients with COVID-19 (n = 60) as compared with healthy controls (n = 17); the number of degranulated MC in lung autopsies was also elevated²⁴² (Table 4). A fourth study reported that lung autopsies from COVID-19 lungs had a 3-fold increase in MC between low- and high-density areas with diffuse alveolar damage (DAD)³⁷⁰ (Table 4). Interestingly, one study reported that MC promotes viral entry of SARS-CoV-2 through the formation of chymase-spike complexes.³⁶²

Infection with SARS-CoV-2 may contribute to neurologic^{239,240,371–375} and psychiatric^{376–382} disorders. Approximately one-half of COVID-19 survivors experience fatigue and other neuropsychiatric symptoms, especially the presence of mental fatigue or brain fog, referred to as Long-COVID.^{383–385} This persistent fatigue is independent of the severity of the initial symptoms.³⁸⁶ Perivascular damage and inflammation^{387–389} has also been reported in the brains of patients with COVID-19,³⁹⁰ but also dysregulation of innate and adaptive immune responses^{391,392} and autoimmunity.³⁷⁵ Symptoms experienced by patients with Long-COVID, especially cognitive impairment,^{287,393} are similar^{394,395} to those present in patients with ME/CFS.^{172,289} Such symptoms are also present in those with MCAS^{396,397} or systemic mastocytosis.³

MC activation was in fact found to be associated with Long-COVID. One study investigated post-acute symptomatic (PASC = 49), post-acute asymptomatic (PAAC = 48) patients and healthy controls (n = 20) and patients with COVID-19 from a previous cohort; levels of chymase, tryptase, and carboxypeptidase A3 were found to be elevated in those with active COVID-19, but only tryptase level remained elevated in PASC but significantly lower than in COVID-19 (Table 4).³⁴⁶ We recently reported that recombinant SARS-CoV-2 spike protein could stimulate cultured human MC to release chymase and tryptase and proinflammatory cytokines, an effect augmented by the cytokine IL-33.^{398,399} We further found that SARS-CoV-2 spike protein could stimulate cultured human microglia to release IL-1 β , IL6, TNF α , and S100B through activation of different receptors.⁴⁰⁰ Obviously one would wonder how MC and microglia may continue to be stimulated past the initial infection. A recent review discussed evidence that the SARS-CoV-2 in its spike protein may persist in certain “reservoirs.”⁴⁰¹ In particular, SARS-CoV-2 spike protein could be detected in patients with Long-COVID for 6 to 12 months.⁴⁰² Other authors reported that the spike protein persisted in CD16+ monocytes for up to 15 months post-infection⁴⁰³ and was associated with sustained CD4+ responses to the spike protein.⁴⁰⁴ Another paper reported that the spike protein may accumulate in the brain.⁴⁰⁵

We recently reported a few common genes involved in the regulation of immune responses in both ME/CFS and COVID-19,⁴⁰⁶ including human leukocyte antigens.^{407,408} We further hypothesized that the spike protein could contribute to not only COVID-19³⁶⁰ but also Long-COVID syndrome,^{359,360,395,409} especially neuro-COVID.³¹

Conclusion

The available evidence indicates that MC can affect the ANS through a variety of actions, such as activation of microglia and

astrocytes, BBB disruption, cardiac pacemaking, circadian clock, coronary artery constriction, HPA axis regulation, neuroinflammation, neurosensitization, and vascular tone control (Table 3), thus potentially contributing to a number of dysautonomias and neuroinflammatory conditions with dysfunctional homeostasis.

However, the presented available evidence of the possible involvement of MC in the ANS is not uniform, often circumstantial, and with many contradicting reports depending on the species and trigger. Moreover, human studies in some of the conditions discussed were based on a few subjects who were not always well defined. Nevertheless, given the many ways MC could be activated and their numerous mediators released, it would make sense to inhibit the activation of MC. Better clinical studies are necessary as is the development of an in vitro disease surrogate system. We are presently working on developing neuroimmune organoids^{410–412} using human-induced pluripotent stem cell–derived MC,⁴¹³ co-cultured with endothelial cells, microglial cells, and neurons using a multi-channel microfluidic model where neuroimmune interactions can be studied under defined conditions that also allow testing of effective inhibitors.

Disclosures

Dr Theoharides is the Scientific Director and shareholder of Algonot, LLC, a company that develops and sells dietary supplements. Dr Theoharides has been awarded the following patents: US 5,855,884; US 6,020,305; US 8,268,365; and EPO 0618796 licensed to Algonot, LLC.

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References

- Gurish MF, Austen KF. Developmental origin and functional specialization of mast cell subsets. *Immunity*. 2012;37(1):25–33.
- Olivera A, Beaven MA, Metcalfe DD. Mast cells signal their importance in health and disease. *J Allergy Clin Immunol*. 2018;142(2):381–393.
- Theoharides TC, Valent P, Akin C. Mast cells, mastocytosis, and related disorders. *N Engl J Med*. 2015;373(2):163–172.
- Valent P, Hartmann K, Bonadonna P, Gülen T, Brockow K, Alvarez-Twose I, et al. Global classification of mast cell activation disorders: an ICD-10-CM-adjusted proposal of the ECNM-AIM consortium. *J Allergy Clin Immunol Pract*. 2022;10(8):1941–1950.
- Theoharides TC, Perlman AI, Twahir A, Kempuraj D. Mast cell activation: beyond histamine and tryptase. *Expert Rev Clin Immunol*. 2023;19(6):639–654.
- Parwaresch MR, Horny HP, Lennert K. Tissue mast cells in health and disease. *Pathol Res Pract*. 1985;179(4–5):439–461.
- Csaba G. Mast cell, the peculiar member of the immune system: a homeostatic aspect. *Acta Microbiol Immunol Hung*. 2015;62(3):207–231.
- Siebenhaar F, Redegeld FA, Bischoff SC, Gibbs BF, Maurer M. Mast cells as drivers of disease and therapeutic targets. *Trends Immunol*. 2018;39(2):151–162.
- Phillips RE, Looareesuwan S, White NJ, Silamut K, Kietinun S, Warrell DA. Quinine pharmacokinetics and toxicity in pregnant and lactating women with falciparum malaria. *Br J Clin Pharmacol*. 1986;21(6):677–683.
- Falduto GH, Pfeiffer A, Luker A, Metcalfe DD, Olivera A. Emerging mechanisms contributing to mast cell-mediated pathophysiology with therapeutic implications. *Pharmacol Ther*. 2021;220:107718.
- Dahlin JS, Maurer M, Metcalfe DD, Pejler G, Sagi-Eisenberg R, Nilsson G. The ingenious mast cell: contemporary insights into mast cell behavior and function. *Allergy*. 2022;77(1):83–99.
- Kolkhir P, Elieh-Ali-Komi D, Metz M, Siebenhaar F, Maurer M. Understanding human mast cells: lesson from therapies for allergic and non-allergic diseases. *Nat Rev Immunol*. 2022;22(5):294–308.
- Levi-Schaffer F, Gibbs BF, Hallgren J, Pucillo C, Redegeld F, Siebenhaar F, et al. Selected recent advances in understanding the role of human mast cells in health and disease. *J Allergy Clin Immunol*. 2022;149(6):1833–1844.
- Sibillano R, Frossi B, Pucillo CE. Mast cell activation: a complex interplay of positive and negative signaling pathways. *Eur J Immunol*. 2014;44(9):2558–2566.
- Gallenga CE, Pandolfi F, Caraffa A, Kritas SK, Ronconi G, Toniato E, et al. Interleukin-1 family cytokines and mast cells: activation and inhibition. *J Biol Regul Homeost Agents*. 2019;33(1):1–6.
- Hakim-Rad K, Metz M, Maurer M. Mast cells: makers and breakers of allergic inflammation. *Curr Opin Allergy Clin Immunol*. 2009;9(5):427–430.
- Theoharides TC, Alysandratos KD, Angelidou A, Delivanis DA, Sismanopoulos N, Zhang B, et al. Mast cells and inflammation. *Biochim Biophys Acta*. 2012;1822(1):21–33.
- Kempuraj D, Ahmed ME, Selvakumar GP, Thangavel R, Dhaliwal AS, Dubova I, et al. Brain injury-mediated neuroinflammatory response and Alzheimer's disease. *Neuroscientist*. 2020;26(2):134–155.
- Galli SJ, Tsai M, Piliponsky AM. The development of allergic inflammation. *Nature*. 2008;454(7203):445–454.
- Mukai K, Tsai M, Saito H, Galli SJ. Mast cells as sources of cytokines, chemokines, and growth factors. *Immunol Rev*. 2018;282(1):121–150.
- Taracanova A, Tsilioni I, Conti P, Norwitz ER, Leeman SE, Theoharides TC. Substance P and IL-33 administered together stimulate a marked secretion of IL-1β from human mast cells, inhibited by methoxyluteolin. *Proc Natl Acad Sci U S A*. 2018;115(40):E9381–E9390.
- Kandere-Grzybowska K, Letourneau R, Kempuraj D, Donelan J, Poplawski S, Boucher W, et al. IL-1 induces vesicular secretion of IL-6 without degranulation from human mast cells. *J Immunol*. 2003;171(9):4830–4836.
- Petra AI, Tsilioni I, Taracanova A, Katsarou-Katsari A, Theoharides TC. Interleukin 33 and interleukin 4 regulate interleukin 31 gene expression and secretion from human laboratory of allergic diseases 2 mast cells stimulated by substance P and/or immunoglobulin E. *Allergy Asthma Proc*. 2018;39(2):153–160.
- Taracanova A, Alevizos M, Karagkouni A, Weng Z, Norwitz E, Conti P, et al. SP and IL-33 together markedly enhance TNF synthesis and secretion from human mast cells mediated by the interaction of their receptors. *Proc Natl Acad Sci U S A*. 2017;114(20):E4002–E4009.
- Horny HP, Valent P. Diagnosis of mastocytosis: general histopathological aspects, morphological criteria, and immunohistochemical findings. *Leuk Res*. 2001;25(7):543–551.
- Valent P, Hartmann K, Bonadonna P, Niedoszytko M, Triggiani M, Arock M, et al. Mast cell activation syndromes: collegium internationale Allergologicum update 2022. *Int Arch Allergy Immunol*. 2022;183(7):693–705.
- Killock D. A new standard for mastocytosis. *Nat Rev Clin Oncol*. 2022;19(2):71.
- Shomali W, Gotlib J. Response criteria in advanced systemic mastocytosis: evolution in the era of KIT inhibitors. *Int J Mol Sci*. 2021;22(6):2983.
- Li Z. New insights into the pathogenesis of systemic mastocytosis. *Int J Mol Sci*. 2021;22(9):4900.
- Theoharides TC, Stewart JM, Hatzigelaki E, Kolaitis G. Brain “fog,” inflammation and obesity: key aspects of neuropsychiatric disorders improved by luteolin. *Front Neurosci*. 2015;9:225.
- Theoharides TC, Kempuraj D. Role of SARS-CoV-2 spike-protein-induced activation of microglia and mast cells in the pathogenesis of neuro-COVID. *Cells*. 2023;12(5):688.
- Moura DS, Sultan S, Georgin-Lavialle S, Barete S, Lortholary O, Gaillard R, et al. Evidence for cognitive impairment in mastocytosis: prevalence, features and correlations to depression. *PLoS One*. 2012;7(6):e39468.
- Afrin LB, Pohlau D, Raithel M, Haenisch B, Dumoulin FL, Homann J, et al. Mast cell activation disease: an underappreciated cause of neurologic and psychiatric symptoms and diseases. *Brain Behav Immun*. 2015;50:314–321.
- Spolak-Bobryk N, Romantowski J, Kujawska-Danecka H, Niedoszytko M. Mastocytosis patients' cognitive dysfunctions correlate with the presence of spindle-shaped mast cells in bone marrow. *Clin Transl Allergy*. 2022;12(1):e12093.
- Boddaert N, Salvador A, Chandesris MO, Lemaître H, Grévent D, Gauthier C, et al. Neuroimaging evidence of brain abnormalities in mastocytosis. *Transl Psychiatry*. 2017;7(8):e1197.
- Theoharides TC, Papaliadis D, Tagen M, Konstantinidou A, Kempuraj D, Clemons A. Chronic fatigue syndrome, mast cells, and tricyclic antidepressants. *J Clin Psychopharmacol*. 2005;25(6):515–520.
- Theoharides TC, Donelan JM, Papadopolou N, Cao J, Kempuraj D, Conti P. Mast cells as targets of corticotropin-releasing factor and related peptides. *Trends Pharmacol Sci*. 2004;25(11):563–568.
- Gordon JR, Galli SJ. Mast cells as a source of both preformed and immunologically inducible TNF-α/cachectin. *Nature*. 1990;346(6281):274–276.
- Zhang B, Alysandratos KD, Angelidou A, Asadi S, Sismanopoulos N, Delivanis DA, et al. Human mast cell degranulation and preformed TNF secretion require mitochondrial translocation to exocytosis sites: relevance to atopic dermatitis. *J Allergy Clin Immunol*. 2011;127(6):1522–1531.e8.
- Zhang B, Asadi S, Weng Z, Sismanopoulos N, Theoharides TC. Stimulated human mast cells secrete mitochondrial components that have autocrine and paracrine inflammatory actions. *PLoS One*. 2012;7(12):e49767.
- Collins LV, Hajizadeh S, Holme E, Jonsson IM, Tarkowski A. Endogenously oxidized mitochondrial DNA induces in vivo and in vitro inflammatory responses. *J Leukoc Biol*. 2004;75(6):995–1000.
- Sun S, Sursal T, Adibnia Y, Zhao C, Zheng Y, Li H, et al. Mitochondrial DAMPs increase endothelial permeability through neutrophil dependent and independent pathways. *PLoS One*. 2013;8(3):e59989.
- Bawazeer MA, Theoharides TC. IL-33 stimulates human mast cell release of CCL5 and CCL2 via MAPK and NF-κappaB, inhibited by methoxyluteolin. *Eur J Pharmacol*. 2019;865:172760.

44. Gagari E, Tsai M, Lantz CS, Fox LG, Galli SJ. Differential release of mast cell interleukin-6 via c-kit. *Blood*. 1997;89(8):2654–2663.
45. Theoharides TC, Boucher W, Spear K. Serum interleukin-6 reflects disease severity and osteoporosis in mastocytosis patients. *Int Arch Allergy Immunol*. 2002;128(4):344–350.
46. Brockow K, Akin C, Huber M, Metcalfe DD. IL-6 levels predict disease variant and extent of organ involvement in patients with mastocytosis. *Clin Immunol*. 2005;115(2):216–223.
47. Mayado A, Teodosio C, Garcia-Montero AC, Matito A, Rodriguez-Caballero A, Morgado JM, et al. Increased IL6 plasma levels in indolent systemic mastocytosis patients are associated with high risk of disease progression. *Leukemia*. 2016;30(1):124–130.
48. Kaur D, Gomez E, Doe C, Berair R, Woodman L, Saunders R, et al. IL-33 drives airway hyper-responsiveness through IL-13-mediated mast cell: airway smooth muscle crosstalk. *Allergy*. 2015;70(5):556–567.
49. Abraham SN, St John AL. Mast cell-orchestrated immunity to pathogens. *Nat Rev Immunol*. 2010;10(6):440–452.
50. Song ST, Wu ML, Zhang HJ, Su X, Wang JH. Mast cell activation triggered by retrovirus promotes acute viral infection. *Front Microbiol*. 2022;13: 798660.
51. Haque TT, Frischmeyer-Guerrero PA. The role of TGF β and other cytokines in regulating mast cell functions in allergic inflammation. *Int J Mol Sci*. 2022;23(18):10864.
52. Gross AR, Theoharides TC. Chondroitin sulfate inhibits secretion of TNF and CXCL8 from human mast cells stimulated by IL-33. *BioFactors*. 2019;45(1):49–61.
53. Lauritano D, Mastrangelo F, D'Ovidio C, Ronconi G, Caraffa A, Gallenga CE, et al. Activation of mast cells by neuropeptides: the role of pro-inflammatory and anti-inflammatory cytokines. *Int J Mol Sci*. 2023;24(5):4811.
54. Yu Y, Blokhuis BR, Garssen J, Redegeld FA. Non-IgE mediated mast cell activation. *Eur J Pharmacol*. 2016;778:33–43.
55. Theoharides TC, Leeman SE. Effect of IL-33 on de novo synthesized mediators from human mast cells. *J Allergy Clin Immunol*. 2019;143(1):451.
56. Theoharides TC. Neuroendocrinology of mast cells: challenges and controversies. *Exp Dermatol*. 2017;26(9):751–759.
57. Theoharides TC, Zhang B, Kempuraj D, Tegen M, Vasiadi M, Angelidou A, et al. IL-33 augments substance P-induced VEGF secretion from human mast cells and is increased in psoriatic skin. *Proc Natl Acad Sci U S A*. 2010;107(9):4448–4453.
58. Sumpter TL, Ho CH, Pleet AR, Tkacheva OA, Shufesky WJ, Rojas-Canales DM, et al. Autocrine hemokinin-1 functions as an endogenous adjuvant for IgE-mediated mast cell inflammatory responses. *J Allergy Clin Immunol*. 2015;135(4):1019–1030.e8.
59. Mashaghi A, Marmalidou A, Tehrani M, Grace PM, Pothoulakis C, Dana R. Neuropeptide substance P and the immune response. *Cell Mol Life Sci*. 2016;73(22):4249–4264.
60. O'Connor TM, O'Connell J, O'Brien DI, Goode T, Bredin CP, Shanahan F. The role of substance P in inflammatory disease. *J Cell Physiol*. 2004;201(2):167–180.
61. Hokfelt T, Pernow B, Wahren J. Substance P: a pioneer amongst neuropeptides. *J Intern Med*. 2001;249(1):27–40.
62. Douglas SD, Leeman SE. Neurokinin-1 receptor: functional significance in the immune system in reference to selected infections and inflammation. *Ann N Y Acad Sci*. 2011;1217:83–95.
63. Suvas S. Role of substance P neuropeptide in inflammation, wound healing, and tissue homeostasis. *J Immunol*. 2017;199(5):1543–1552.
64. Saluja R, Khan M, Church MK, Maurer M. The role of IL-33 and mast cells in allergy and inflammation. *Clin Transl Allergy*. 2015;5:33.
65. Theoharides TC, Kempuraj D, Tegen M, Conti P, Kalogeromitos D. Differential release of mast cell mediators and the pathogenesis of inflammation. *Immunol Rev*. 2007;217:65–78.
66. Theoharides TC, Tsilioni I, Bawazeer M. Mast cells, neuroinflammation and pain in fibromyalgia syndrome. *Front Cell Neurosci*. 2019;13:353.
67. Xu H, Shi X, Li X, Zou J, Zhou C, Liu W, et al. Neurotransmitter and neuropeptide regulation of mast cell function: a systematic review. *J Neuroinflammation*. 2020;17(1):356.
68. Levi-Montalcini R, Skaper SD, Dal Toso R, Petrelli L, Leon A. Nerve growth factor: from neurotrophin to neurokinine. *Trends Neurosci*. 1996;19(11):514–520.
69. Donelan J, Boucher W, Papadopoulos N, Lytinas M, Papaliodis D, Dobner P, et al. Corticotropin-releasing hormone induces skin vascular permeability through a neurotensin-dependent process. *Proc Natl Acad Sci U S A*. 2006;103(20):7759–7764.
70. Theoharides TC, Betchaku T, Douglas WW. Somatostatin-induced histamine secretion in mast cells. Characterization of the effect. *Eur J Pharmacol*. 1981;69(2):127–137.
71. Theoharides TC, Douglas WW. Mast cell histamine secretion in response to somatostatin analogues: structural considerations. *Eur J Pharmacol*. 1981;73(2-3):131–136.
72. Kumar M, Duraisamy K, Chow BK. Unlocking the non-IgE-mediated pseudo-allergic reaction puzzle with Mas-related G-protein coupled receptor member X2 (MRGPRX2). *Cells*. 2021;10(5):1033.
73. Chompunud Na Ayudhya C, Ali H. Mas-related G protein-coupled receptor-X2 and its role in non-immunoglobulin E-mediated drug hypersensitivity. *Immunol Allergy Clin North Am*. 2022;42(2):269–284.
74. Roy S, Chompunud Na Ayudhya C, Thapaliya M, Deepak V, Ali H. Multifaceted MRGPRX2: new insight into the role of mast cells in health and disease. *J Allergy Clin Immunol*. 2021;148(2):293–308.
75. Xu H, Bin NR, Sugita S. Diverse exocytic pathways for mast cell mediators. *Biochem Soc Trans*. 2018;46(2):235–247.
76. Gilfillan AM, Tkaczuk C. Integrated signalling pathways for mast-cell activation. *Nat Rev Immunol*. 2006;6(3):218–230.
77. Gaudenzio N, Sibillano R, Marichal T, Starkl P, Reber LL, Cenac N, et al. Different activation signals induce distinct mast cell degranulation strategies. *J Clin Invest*. 2016;126(10):3981–3998.
78. Theoharides TC. The impact of psychological stress on mast cells. *Ann Allergy Asthma Immunol*. 2020;125(4):388–392.
79. Kleij HP, Bienenstock J. Significance of conversation between mast cells and nerves. *Allergy Asthma Clin Immunol*. 2005;1(2):65–80.
80. Rozniecki JJ, Dimitriadou V, Lambricht-Hall M, Pang X, Theoharides TC. Morphological and functional demonstration of rat dura mater mast cell-neuron interactions in vitro and in vivo. *Brain Res*. 1999;849(1-2):1–15.
81. Weng Z, Zhang B, Tsilioni I, Theoharides TC. Nanotube formation: a rapid form of "alarm signaling"? *Clin Ther*. 2016;38(5):1066–1072.
82. Heath D, Lowe P, Smith P. Mast cells in the human carotid body. *J Clin Pathol*. 1987;40(1):9–12.
83. Kounis NG, Hahalis G, Theoharides TC. Coronary stents, hypersensitivity reactions, and the Kounis syndrome. *J Interv Cardiol*. 2007;20(5):314–323.
84. Patella V, Marino I, Lamparter B, Arbustini E, Adt M, Marone G. Human heart mast cells. Isolation, purification, ultrastructure, and immunologic characterization. *J Immunol*. 1995;154(6):2855–2865.
85. Poto R, Patella V, Criscuolo G, Marone G, Coscioni E, Varricchi G. Autoantibodies to IgE can induce the release of proinflammatory and vasoactive mediators from human cardiac mast cells. *Clin Exp Med*. 2023;23(4):1265–1276.
86. Forsythe P. The parasympathetic nervous system as a regulator of mast cell function. *Methods Mol Biol*. 2015;1220:141–154.
87. Stead RH, Colley EC, Wang B, Partosoedarmo E, Lin J, Stanis A, et al. Vagal influences over mast cells. *Auton Neurosci*. 2006;125(1-2):53–61.
88. Wessler I, Kilbinger H, Bittiger F, Kirkpatrick CJ. The biological role of non-neuronal acetylcholine in plants and humans. *Jpn J Pharmacol*. 2001;85(1):2–10.
89. Wessler I, Reinheimer T, Kilbinger H, Bittiger F, Kirkpatrick CJ, Saloga J, et al. Increased acetylcholine levels in skin biopsies of patients with atopic dermatitis. *Life Sci*. 2003;72(18-19):2169–2172.
90. Tokura Y. Direct and indirect action modes of acetylcholine in cholinergic urticaria. *Allergol Int*. 2021;70(1):39–44.
91. Fantozzi R, Masini E, Blandina P, Mannaioni PF, Bani-Sacchi T. Release of histamine from rat mast cells by acetylcholine. *Nature*. 1978;273(5662):473–474.
92. Erjavec F, Ferjan I. The mechanism of histamine release induced by pilocarpine from different tissues: studies on rat peritoneal mast cells. *Agents Actions*. 1994;41(special no):C32–C33.
93. Kazimierzczak W, Ada mas B, Maslinski C. Failure of acetylcholine to release histamine from rat mast cells. *Agents Actions*. 1980;10(1 Pt 2):1–3.
94. Erjavec F, Iskra M. The interaction of cholinomimetics, peptides and compounds 48/80 on histamine secretion from the mast cell. *Agents Actions*. 1984;14(3-4):373–375.
95. Hanna CJ, Johnston ME, Schellenberg RR. Evaluation of the action of cholinergic agonists and cyclic guanosine monophosphate (cGMP) on histamine release from purified rat mast cells. *Immunopharmacology*. 1981;3(3):233–240.
96. Reinheimer T, Baumgartner D, Hohle KD, Racke K, Wessler I. Acetylcholine via muscarinic receptors inhibits histamine release from human isolated bronchi. *Am J Respir Crit Care Med*. 1997;156(2 Pt 1):389–395.
97. Kageyama-Yahara N, Suehiro Y, Yamamoto T, Kadowaki M. IgE-induced degranulation of mucosal mast cells is negatively regulated via nicotinic acetylcholine receptors. *Biochem Biophys Res Commun*. 2008;377(1):321–325.
98. Ertle CM, Rommel FR, Tumala S, Moriwaki Y, Klein J, Kruse J, et al. New pathways for the skin's stress response: the cholinergic neuropeptide SLURP-1 can activate mast cells and alter cytokine production in mice. *Front Immunol*. 2021;12: 631881.
99. Yamamoto T, Kodama T, Lee J, Utsunomiya N, Hayashi S, Sakamoto H, et al. Anti-allergic role of cholinergic neuronal pathway via $\alpha 7$ nicotinic ACh receptors on mucosal mast cells in a murine food allergy model. *PLoS One*. 2014;9(1):e85888.
100. Mishra NC, Rir-sima-ah J, Boyd RT, Singh SP, Gundavarapu S, Langley RJ, et al. Nicotine inhibits Fc epsilon RI-induced cysteinyl leukotrienes and cytokine production without affecting mast cell degranulation through alpha 7/alpha 9/alpha 10-nicotinic receptors. *J Immunol*. 2010;185(1):588–596.
101. Masini E, Fantozzi R, Blandina P, et al. Presence of functionally active beta-adrenoceptors in rat mast cells. Correlation between (–)[3H]-dihydroalprenolol binding and inhibition of histamine release. *Naunyn Schmiedeberg Arch Pharmacol*. 1982;321(3):171–176.
102. Pham L, Baiocchi L, Kennedy L, Sato K, Meadows V, Meng F, et al. The interplay between mast cells, pineal gland, and circadian rhythm: links between histamine, melatonin, and inflammatory mediators. *J Pineal Res*. 2021;70(2):e12699.
103. Wang X, Reece SP, Van Scott MR, Brown JM. A circadian clock in murine bone marrow-derived mast cells modulates IgE-dependent activation in vitro. *Brain Behav Immun*. 2011;25(1):127–134.
104. Kawauchi T, Ishimaru K, Nakamura Y, Nakano N, Hara M, Ogawa H, et al. Clock-dependent temporal regulation of IL-33/ST2-mediated mast cell response. *Allergol Int*. 2017;66(3):472–478.
105. Christ P, Sowa AS, Froy O, Lorentz A. The circadian clock drives mast cell functions in allergic reactions. *Front Immunol*. 2018;9:1526.
106. Kvetnoy IM. Extrapineal melatonin: location and role within diffuse neuroendocrine system. *Histochem J*. 1999;31(1):1–12.
107. Bao C, Chen O, Sheng H, Zhang J, Luo Y, Hayes BW, et al. A mast cell-thermoregulatory neuron circuit axis regulates hypothermia in anaphylaxis. *Sci Immunol*. 2023;8(81):ead9417.

108. Parsons IT, Stacey MJ, Faconti L, Hill N, O'Hara J, Walter E, et al. Histamine, mast cell tryptase and post-exercise hypotension in healthy and collapsed marathon runners. *Eur J Appl Physiol*. 2021;121(5):1451–1459.
109. Novak P, Giannetti MP, Weller E, Hamilton MJ, Castells M. Mast cell disorders are associated with decreased cerebral blood flow and small fiber neuropathy. *Ann Allergy Asthma Immunol*. 2022;128(3):299–306.e1.
110. Theoharides TC. Mast cells: the immune gate to the brain. *Life Sci*. 1990;46(9):607–617.
111. Traina G. Mast cells in the brain – old cells, new target. *J Integr Neurosci*. 2017;16(s1):S69–S83.
112. Polyzoidis S, Koletsis T, Panagiotidou S, Ashkan K, Theoharides TC. Mast cells in meningiomas and brain inflammation. *J Neuroinflammation*. 2015;12:170.
113. Pang X, Letourneau R, Rozniecki JJ, Wang L, Theoharides TC. Definitive characterization of rat hypothalamic mast cells. *Neuroscience*. 1996;73(3):889–902.
114. Theoharides TC, Konstantinidou AD. Corticotropin-releasing hormone and the blood-brain-barrier. *Front Biosci*. 2007;12:1615–1628.
115. Colonna M, Butovsky O. Microglia function in the central nervous system during health and neurodegeneration. *Annu Rev Immunol*. 2017;35:441–468.
116. Edvinsson L, Cervos-Navarro J, Larsson LI, Owman C, Ronnberg AL. Regional distribution of mast cells containing histamine, dopamine, or 5-hydroxytryptamine in the mammalian brain. *Neurology*. 1977;27(9):878–883.
117. Olsson Y. Mast cells in the nervous system. *Int Rev Cytol*. 1968;24:27–70.
118. Olsson Y. Mast cells in plaques of multiple sclerosis. *Acta Neurol Scand*. 1974;50(5):611–618.
119. Kruger PG, Bo L, Myhr KM, Karlsen AE, Taule A, Nyland HI, et al. Mast cells and multiple sclerosis: a light and electron microscopic study of mast cells in multiple sclerosis emphasizing staining procedures. *Acta Neurol Scand*. 1990;81(1):31–36.
120. Toms R, Weiner HL, Johnson D. Identification of IgE-positive cells and mast cells in frozen sections of multiple sclerosis brains. *J Neuroimmunol*. 1990;30(2–3):169–177.
121. Rozniecki JJ, Hauser SL, Stein M, Lincoln R, Theoharides TC. Elevated mast cell tryptase in cerebrospinal fluid of multiple sclerosis patients. *Ann Neurol*. 1995;37(1):63–66.
122. Smith JH, Snyder MR, Weiler CR, Cutrer FM, Butterfield JH. Lack of evidence for intrathecal tryptase synthesis in patients with systemic mastocytosis. *J Allergy Clin Immunol*. 2013;132(4):996–997.
123. Matsumoto I, Inoue Y, Shimada T, Aikawa T. Brain mast cells act as an immune gate to the hypothalamic-pituitary-adrenal axis in dogs. *J Exp Med*. 2001;194(1):71–78.
124. Bugajski AJ, Chlap Z, Gadek-Michalska A, Borycz J, Bugajski J. Degranulation and decrease in histamine levels of thalamic mast cells coincides with corticosterone secretion induced by compound 48/80. *Inflamm Res*. 1995;44(suppl 1):S50–S51.
125. Kalogeromitros D, Syrigou EK, Makris M, Kempuraj D, Stavrianeas NG, Vasiadi M, et al. Nasal provocation of patients with allergic rhinitis and the hypothalamic-pituitary-adrenal axis. *Ann Allergy Asthma Immunol*. 2007;98(3):269–273.
126. Foldes A, Nemethy Z, Szalay O, Kovacs KJ. Anaphylactoid reactions activate hypothalamo-pituitary-adrenocortical axis: comparison with endotoxic reactions. *Brain Res Bull*. 2000;52(6):573–579.
127. Scaccianoce S, Lombardo K, Nicolai R, Affricano D, Angelucci L. Studies on the involvement of histamine in the hypothalamic-pituitary-adrenal axis activation induced by nerve growth factor. *Life Sci*. 2000;67(26):3143–3152.
128. Spath-Schwalbe E, Born J, Schrezenmeier H, Bornstein SR, Stromeyer P, Drechsler S, et al. Interleukin-6 stimulates the hypothalamus-pituitary-adrenocortical axis in man. *J Clin Endocrinol Metab*. 1994;79(4):1212–1214.
129. Kempuraj D, Papadopoulou NG, Lytinas M, Huang M, Kandere-Grzybowska K, Madhappan B, et al. Corticotropin-releasing hormone and its structurally related urocortin are synthesized and secreted by human mast cells. *Endocrinology*. 2004;145(1):43–48.
130. Schilling L, Wahl M. Opening of the blood-brain barrier during cortical superfusion with histamine. *Brain Res*. 1994;653(1–2):289–296.
131. Black KL. Biochemical opening of the blood-brain barrier. *Adv Drug Deliv Rev*. 1995;15(1–3):37–52.
132. Sedeyn JC, Wu H, Hobbs RD, Levin EC, Nagele RG, Venkataraman V. Histamine induces Alzheimer's disease-like blood brain barrier breach and local cellular responses in mouse brain organotypic cultures. *BioMed Res Int*. 2015;2015:937148.
133. Yue J, Tan Y, Huan R, Guo J, Yang S, Deng M, et al. Mast cell activation mediates blood-brain barrier impairment and cognitive dysfunction in septic mice in a histamine-dependent pathway. *Front Immunol*. 2023;14: 1090288.
134. Zhou Q, Wang YW, Ni PF, Chen YN, Dong HQ, Qian YN. Effect of tryptase on mouse brain microvascular endothelial cells via protease-activated receptor 2. *J Neuroinflammation*. 2018;15(1):248.
135. Abbott NJ. Inflammatory mediators and modulation of blood-brain barrier permeability. *Cell Mol Neurobiol*. 2000;20(2):131–147.
136. Pan W, Stone KP, Hsueh H, Manda VK, Zhang Y, Kastin AJ. Cytokine signaling modulates blood-brain barrier function. *Curr Pharm Des*. 2011;17(33):3729–3740.
137. Esposito P, Chandler N, Kandere K, Basu S, Jacobson S, Connolly R, et al. Corticotropin-releasing hormone and brain mast cells regulate blood-brain-barrier permeability induced by acute stress. *J Pharmacol Exp Ther*. 2002;303(3):1061–1066.
138. Sarlus H, Hoglund CO, Karshikoff B, Wang X, Lekander M, Schultzberg M, et al. Allergy influences the inflammatory status of the brain and enhances tau-phosphorylation. *J Cell Mol Med*. 2012;16(10):2401–2412.
139. Banks WA, Kastin AJ, Broadwell RD. Passage of cytokines across the blood-brain barrier. *Neuroimmunomodulation*. 1995;2(4):241–248.
140. Romanovsky AA, Ivanov AI, Karman EK. Blood-borne, albumin-bound prostaglandin E2 may be involved in fever. *Am J Physiol*. 1999;276(6):R1840–R1844.
141. Dimitriadou V, Rouleau A, Trung Tuong MD, Newlands GJ, Miller HR, Luffau G, et al. Functional relationships between sensory nerve fibers and mast cells of dura mater in normal and inflammatory conditions. *Neuroscience*. 1997;77(3):829–839.
142. Paus R, Theoharides TC, Arck PC. Neuroimmunoendocrine circuitry of the 'brain-skin connection'. *Trends Immunol*. 2006;27(1):32–39.
143. Kempuraj D, Ahmed ME, Selvakumar GP, Thangavel R, Raikwar SP, Zaheer SA, et al. Mast cell activation, neuroinflammation, and tight junction protein derangement in acute traumatic brain injury. *Mediators Inflamm*. 2020;2020: 4243953.
144. Theoharides TC. Effect of stress on neuroimmune processes. *Clin Ther*. 2020;42(6):1007–1014.
145. Fiorentino M, Sapone A, Senger S, Camhi SS, Kadzielski SM, Buie TM, et al. Blood-brain barrier and intestinal epithelial barrier alterations in autism spectrum disorders. *Mol Autism*. 2016;7:49.
146. Rozniecki JJ, Sahagian GG, Kempuraj D, Tao K, Jacobson S, Zhang B, et al. Brain metastases of mouse mammary adenocarcinoma is increased by acute stress. *Brain Res*. 2010;1366:204–210.
147. Theoharides TC, Rozniecki JJ, Sahagian G, Jacobson S, Kempuraj D, Conti P, et al. Impact of stress and mast cells on brain metastases. *J Neuroimmunol*. 2008;205(1–2):1–7.
148. Kandere-Grzybowska K, Gheorghe D, Priller J, Esposito P, Huang M, Gerard N, et al. Stress-induced dura vascular permeability does not develop in mast cell-deficient and neurokinin-1 receptor knockout mice. *Brain Res*. 2003;980(2):213–220.
149. Sayed BA, Christy AL, Walker ME, Brown MA. Meningeal mast cells affect early T cell central nervous system infiltration and blood-brain barrier integrity through TNF: a role for neutrophil recruitment? *J Immunol*. 2010;184(12):6891–6900.
150. Wang Y, Sha H, Zhou L, Chen Y, Zhou Q, Dong H, et al. The mast cell is an early activator of lipopolysaccharide-induced neuroinflammation and blood-brain barrier dysfunction in the hippocampus. *Mediators Inflamm*. 2020;2020: 8098439.
151. Kempuraj D, Mentor S, Thangavel R, Ahmed ME, Selvakumar GP, Raikwar SP, et al. Mast cells in stress, pain, blood-brain barrier, neuroinflammation and Alzheimer's disease. *Front Cell Neurosci*. 2019;13:54.
152. Milligan Armstrong A, Porter T, Quek H, White A, Haynes J, Jackaman C, et al. Chronic stress and Alzheimer's disease: the interplay between the hypothalamic-pituitary-adrenal axis, genetics and microglia. *Biol Rev Camb Philos Soc*. 2021;96(5):2209–2228.
153. Alysandratos KD, Asadi S, Angelidou A, Zhang B, Sismanopoulos N, Yang H, et al. Neurotensin and CRH interactions augment human mast cell activation. *PLoS One*. 2012;7(11):e48934.
154. Asadi S, Alysandratos KD, Angelidou A, Miniati A, Sismanopoulos N, Vasiadi M, et al. Substance P (SP) induces expression of functional corticotropin-releasing hormone receptor-1 (CRHR-1) in human mast cells. *J Invest Dermatol*. 2012;132(2):324–329.
155. Okada T, Hirayama Y, Kishi S, Miyayasu K, Hiroi J, Fujii T. Functional neurokinin NK-1 receptor expression in rat peritoneal mast cells. *Inflamm Res*. 1999;48(5):274–279.
156. Nomura H, Shimizume R, Ikegaya Y. Histamine: a key neuromodulator of memory consolidation and retrieval. *Curr Top Behav Neurosci*. 2022;59:329–353.
157. Carthy E, Ellender T. Histamine, neuroinflammation and neurodevelopment: a review. *Front Neurosci*. 2021;15: 680214.
158. Mochizuki T. Histamine as an alert signal in the brain. *Curr Top Behav Neurosci*. 2022;59:413–425.
159. Sadek B, Saad A, Sadeq A, Jalal F, Stark H. Histamine H3 receptor as a potential target for cognitive symptoms in neuropsychiatric diseases. *Behav Brain Res*. 2016;312:415–430.
160. da Silva Meirelles L, Simon D, Regner A. Neurotrauma: the crosstalk between neurotrophins and inflammation in the acutely injured brain. *Int J Mol Sci*. 2017;18(5):1082.
161. Torrealba F, Riveros ME, Contreras M, Valdes JL. Histamine and motivation. *Front Syst Neurosci*. 2012;6:51.
162. Schlicker E, Kathmann M. Role of the histamine H3 receptor in the central nervous system. *Handb Exp Pharmacol*. 2017;241:277–299.
163. Provensi G, Costa A, Izquierdo I, Blandina P, Passani MB. Brain histamine modulates recognition memory: possible implications in major cognitive disorders. *Br J Pharmacol*. 2020;177(3):539–556.
164. Passani MB, Benetti F, Blandina P, Furini CRG, de Carvalho Myskiw J, Izquierdo I. Histamine regulates memory consolidation. *Neurobiol Learn Mem*. 2017;145:1–6.
165. Burgess CR. Histamine and orexin in the control of arousal, locomotion, and motivation. *J Neurosci*. 2010;30(8):2810–2811.
166. da Silva CEK, Furini CR, Benetti F, Monteiro Sda C, Izquierdo I. The role of histamine receptors in the consolidation of object recognition memory. *Neurobiol Learn Mem*. 2013;103:64–71.
167. Zhou L, Chen L, Li X, Li T, Dong Z, Wang YT. Food allergy induces alteration in brain inflammatory status and cognitive impairments. *Behav Brain Res*. 2019;364:374–382.
168. Zhang X, Dong H, Li N, Zhang S, Sun J, Zhang S, et al. Activated brain mast cells contribute to postoperative cognitive dysfunction by evoking microglia activation and neuronal apoptosis. *J Neuroinflammation*. 2016;13(1):127.
169. Zhang X, Wang Y, Dong H, Xu Y, Zhang S. Induction of microglial activation by mediators released from mast cells. *Cell Physiol Biochem*. 2016;38(4):1520–1531.
170. Shaik-Dasthagirisheh YB, Conti P. The role of mast cells in Alzheimer's disease. *Adv Clin Exp Med*. 2016;25(4):781–787.
171. Hansen DV, Hanson JE, Sheng M. Microglia in Alzheimer's disease. *J Cell Biol*. 2018;217(2):459–472.
172. Hatzigelaki E, Adamaki M, Tsilioni I, Dimitriadis G, Theoharides TC. Myalgic encephalomyelitis/chronic fatigue syndrome-metabolic disease or disturbed

- homeostasis due to focal inflammation in the hypothalamus? *J Pharmacol Exp Ther.* 2018;367(1):155–167.
173. Oken D. Fifty years and still going strong!. *Psychosom Med.* 1989;51(6):595–596.
 174. Zhang XJ, Lv MM, Zhu XQ, Tian LY, Li JJ, Shao YP, et al. Microglia M1/M2 polarization contributes to electromagnetic pulse-induced brain injury. *J Biol Regul Homeost Agents.* 2019;33(4):1051–1062.
 175. Liberman AC, Trias E, da Silva Chagas L, Trindade P, Dos Santos Pereira M, Refojo D, et al. Neuroimmune and inflammatory signals in complex disorders of the central nervous system. *Neuroimmunomodulation.* 2018;25(5-6):246–270.
 176. Bachiller S, Jimenez-Ferrer I, Paulus A, Yang Y, Swanberg M, Deierborg T, et al. Microglia in neurological diseases: a road map to brain-disease dependent-inflammatory response. *Front Cell Neurosci.* 2018;12:488.
 177. Subhramanyam CS, Wang C, Hu Q, Dheen ST. Microglia-mediated neuroinflammation in neurodegenerative diseases. *Semin Cell Dev Biol.* 2019;94:112–120.
 178. Perry VH, Nicoll JA, Holmes C. Microglia in neurodegenerative disease. *Nat Rev Neurol.* 2010;6(4):193–201.
 179. Ransohoff RM. How neuroinflammation contributes to neurodegeneration. *Science.* 2016;353(6301):777–783.
 180. Hickman S, Izzy S, Sen P, Morsettt L, El Khoury J. Microglia in neurodegeneration. *Nat Neurosci.* 2018;21(10):1359–1369.
 181. Angelova DM, Brown DR. Microglia and the aging brain: are senescent microglia the key to neurodegeneration? *J Neurochem.* 2019;151(6):676–688.
 182. Luo XG, Ding JQ, Chen SD. Microglia in the aging brain: relevance to neurodegeneration. *Mol Neurodegener.* 2010;5:12.
 183. Nordengen K, Kirsebom BE, Henjum K, Selnes P, Gisladóttir B, Wettergreen M, et al. Glial activation and inflammation along the Alzheimer's disease continuum. *J Neuroinflammation.* 2019;16(1):46.
 184. Paouri E, Georgopoulos S. Systemic and CNS inflammation crosstalk: implications for Alzheimer's disease. *Curr Alzheimer Res.* 2019;16(6):559–574.
 185. Jack CS, Arbour N, Manusow J, Montgrain V, Blain M, McCrea E, et al. TLR signaling tailors innate immune responses in human microglia and astrocytes. *J Immunol.* 2005;175(7):4320–4330.
 186. Shen Z, Bao X, Wang R. Clinical PET imaging of microglial activation: implications for microglial therapeutics in Alzheimer's disease. *Front Aging Neurosci.* 2018;10:314.
 187. Liu B, Hong JS. Role of microglia in inflammation-mediated neurodegenerative diseases: mechanisms and strategies for therapeutic intervention. *J Pharmacol Exp Ther.* 2003;304(1):1–7.
 188. Jha MK, Jo M, Kim JH, Suk K. Microglia-astrocyte crosstalk: an intimate molecular conversation. *Neuroscientist.* 2019;25(3):227–240.
 189. Xu L, He D, Bai Y. Microglia-mediated inflammation and neurodegenerative disease. *Mol Neurobiol.* 2016;53(10):6709–6715.
 190. Ho GJ, Drego R, Hakimian E, Masliah E. Mechanisms of cell signaling and inflammation in Alzheimer's disease. *Curr Drug Targets Inflamm Allergy.* 2005;4(2):247–256.
 191. El Khoury JB, Moore KJ, Means TK, Leung J, Terada K, Toft M, et al. CD36 mediates the innate host response to beta-amyloid. *J Exp Med.* 2003;197(12):1657–1666.
 192. Stewart CR, Stuart LM, Wilkinson K, van Gils JM, Deng J, Halle A, et al. CD36 ligands promote sterile inflammation through assembly of a toll-like receptor 4 and 6 heterodimer. *Nat Immunol.* 2010;11(2):155–161.
 193. Martin S, Dicu E, Vincent JP, Mazella J. Neutensin and the neutensin receptor-3 in microglial cells. *J Neurosci Res.* 2005;81(3):322–326.
 194. Patel AB, Tsilioni I, Leeman SE, Theoharides TC. Neutensin stimulates sortilin and mTOR in human microglia inhibitable by methoxyluteolin, a potential therapeutic target for autism. *Proc Natl Acad Sci U S A.* 2016;113(45):E7049–E7058.
 195. Wang W, Ji P, Riopelle RJ, Dow KE. Functional expression of corticotropin-releasing hormone (CRH) receptor 1 in cultured rat microglia. *J Neurochem.* 2002;80(2):287–294.
 196. Niraula A, Sheridan JF, Godbout JP. Microglia priming with aging and stress. *Neuropharmacology.* 2017;42(1):318–333.
 197. Carrier M, Simoncicova E, St-Pierre MK, McKee C, Tremblay ME. Psychological stress as a risk factor for accelerated cellular aging and cognitive decline: the involvement of microglia-neuron crosstalk. *Front Mol Neurosci.* 2021;14:749737.
 198. Hendriksen E, van Bergeijk D, Oosting RS, Redegeld FA. Mast cells in neuroinflammation and brain disorders. *Neurosci Biobehav Rev.* 2017;79:119–133.
 199. Sandhu JK, Kulka M. Decoding mast cell-microglia communication in neurodegenerative diseases. *Int J Mol Sci.* 2021;22(3):1093.
 200. Skaper SD, Facci L, Giusti P. Neuroinflammation, microglia and mast cells in the pathophysiology of neurocognitive disorders: a review. *CNS Neurol Disord Drug Targets.* 2014;13(10):1654–1666.
 201. Skaper SD, Facci L, Zusso M, Giusti P. Neuroinflammation, mast cells, and glia: dangerous liaisons. *Neuroscientist.* 2017;23(5):478–498.
 202. Skaper SD, Giusti P, Facci L. Microglia and mast cells: two tracks on the road to neuroinflammation. *FASEB J.* 2012;26(8):3103–3117.
 203. Selvakumar GP, Ahmed ME, Thangavel R, Kempuraj D, Dubova I, Raikwar SP, et al. A role for glia maturation factor dependent activation of mast cells and microglia in MPTP induced dopamine loss and behavioural deficits in mice. *Brain Behav Immun.* 2020;87:429–443.
 204. Kempuraj D, Selvakumar GP, Zaheer S, Thangavel R, Ahmed ME, Raikwar S, et al. Cross-talk between glia, neurons and mast cells in neuroinflammation associated with Parkinson's disease. *J Neuroimmune Pharmacol.* 2018;13(1):100–112.
 205. Dong H, Zhang X, Wang Y, Zhou X, Qian Y, Zhang S. Suppression of brain mast cells degranulation inhibits microglial activation and central nervous system inflammation. *Mol Neurobiol.* 2017;54(2):997–1007.
 206. Theoharides TC. Ways to Address Perinatal mast cell Activation and Focal Brain Inflammation, including Response to SARS-CoV-2, in Autism Spectrum Disorder. *J Pers Med.* 2021;11(9):860.
 207. Zhang W, Zhang X, Zhang Y, Qu C, Zhou X, Zhang S. Histamine induces microglia activation and the release of proinflammatory mediators in rat brain via H1R or H4R. *J Neuroimmune Pharmacol.* 2020;15(2):280–291.
 208. Zhang S, Zeng X, Yang H, Hu G, He S. Mast cell tryptase induces microglia activation via protease-activated receptor 2 signaling. *Cell Physiol Biochem.* 2012;29(5-6):931–940.
 209. Kempuraj D, Thangavel R, Selvakumar GP, Ahmed ME, Zaheer S, Raikwar SP, et al. Mast cell proteases activate astrocytes and glia-neurons and release interleukin-33 by activating p38 and ERK1/2 MAPKs and NF- κ B. *Mol Neurobiol.* 2019;56(3):1681–1693.
 210. Dong H, Wang Y, Zhang X, Zhang X, Qian Y, Ding H, et al. Stabilization of brain mast cells alleviates LPS-induced neuroinflammation by inhibiting microglia activation. *Front Cell Neurosci.* 2019;13:191.
 211. Bonamichi-Santos R, Yoshimi-Kanamori K, Giavina-Bianchi P, Aun MV. Association of postural tachycardia syndrome and Ehlers-Danlos syndrome with mast cell activation disorders. *Immunol Allergy Clin North Am.* 2018;38(3):497–504.
 212. Wang E, Ganti T, Vaou E, Hohler A. The relationship between mast cell activation syndrome, postural tachycardia syndrome, and Ehlers-Danlos syndrome. *Allergy Asthma Proc.* 2021;42(3):243–246.
 213. Seneviratne SL, Maitland A, Afrin L. Mast cell disorders in Ehlers-Danlos syndrome. *Am J Med Genet C.* 2017;175(1):226–236.
 214. Afrin LB. Some cases of hypermobile Ehlers-Danlos syndrome may be rooted in mast cell activation syndrome. *Am J Med Genet C.* 2021;187(4):466–472.
 215. Shibao C, Arzubigaa C, Roberts 2nd LJ, Raj S, Black B, Harris P, et al. Hyperadrenergic postural tachycardia syndrome in mast cell activation disorders. *Hypertension.* 2005;45(3):385–390.
 216. Cheung I, Vadas P. A new disease cluster: mast cell activation syndrome, postural orthostatic tachycardia syndrome, and Ehlers-Danlos syndrome. *J Allergy Clin Immunol.* 2015;135(2):AB65.
 217. Song B, Yeh P, Harrell J. Systemic manifestations of Ehlers-Danlos syndrome. *Proc (Bayl Univ Med Cent).* 2020;34(1):49–53.
 218. Morgan AW, Pearson SB, Davies S, Gooi HC, Bird HA. Asthma and airways collapse in two heritable disorders of connective tissue. *Ann Rheum Dis.* 2007;66(10):1369–1373.
 219. Huang KZ, Dellon ES. Increased prevalence of autonomic dysfunction due to postural orthostatic tachycardia syndrome in patients with eosinophilic gastrointestinal disorders. *J Gastrointest Liver Dis.* 2019;28(1):47–51.
 220. Theoharides TC, Tsilioni I, Patel AB, Doyle R. Atopic diseases and inflammation of the brain in the pathogenesis of autism spectrum disorders. *Transl Psychiatry.* 2016;6(6):e844.
 221. Jyonouchi H. Autism spectrum disorders and allergy: observation from a pediatric allergy/immunology clinic. *Expert Rev Clin Immunol.* 2010;6(3):397–411.
 222. Magalhaes ES, Pinto-Mariz F, Bastos-Pinto S, Pontes AT, Prado EA, deAzevedo LC. Immune allergic response in Asperger syndrome. *J Neuroimmunol.* 2009;216(1-2):108–112.
 223. Lyall K, Van de Water J, Ashwood P, Hertz-Picciotto I. Asthma and allergies with autism spectrum disorders: results from the CHARGE study. *Autism Res.* 2015;8(5):567–574.
 224. Fombonne E. Epidemiology of pervasive developmental disorders. *Pediatr Res.* 2009;65(6):591–598.
 225. Jyonouchi H. Food allergy and autism spectrum disorders: is there a link? *Curr Allergy Asthma Rep.* 2009;9(3):194–201.
 226. Li H, Liu H, Chen X, Zhang J, Tong G, Sun Y. Association of food hypersensitivity in children with the risk of autism spectrum disorder: a meta-analysis. *Eur J Pediatr.* 2021;180(4):999–1008.
 227. Xue M, Brimacombe M, Chaaban J, Zimmerman-Bier B, Wagner GC. Autism spectrum disorders: concurrent clinical disorders. *J Child Neurol.* 2008;23(1):6–13.
 228. Kotey S, Ertel K, Whitcomb B. Co-occurrence of autism and asthma in a nationally representative sample of children in the United States. *J Autism Dev Disord.* 2014;44(12):3083–3088.
 229. Zaitis M, Mizoguchi T, Morita S, Kawasaki S, Iwanaga A, Matsuo M. Developmental disorders in school children are related to allergic diseases. *Pediatr Int.* 2022;64(1):e15358.
 230. Billeci L, Tonacci A, Tartarico G, Ruta L, Pioggia G, Gangemi S. Association between atopic dermatitis and autism spectrum disorders: a systematic review. *Am J Clin Dermatol.* 2015;16(5):371–388.
 231. Wan J, Shin DB, Syed MN, Abuabara K, Lemeshow AR, Gelfand JM. Atopic dermatitis and risk of major neuropsychiatric disorders in children: a population-based cohort study. *J Eur Acad Dermatol Venereol.* 2023;37(1):114–122.
 232. Nemet S, Asher I, Voles I, Baevsky T, Shogher Z. Early childhood allergy linked with development of attention deficit hyperactivity disorder and autism spectrum disorder. *Pediatr Allergy Immunol.* 2022;33(6). <https://doi.org/10.1111/pai.13819>.
 233. Gong T, Lundholm C, Lundstrom S, Kuja-Halkola R, Taylor MJ, Almqvist C. Understanding the relationship between asthma and autism spectrum disorder: a population-based family and twin study. *Psychol Med.* 2022;53(7):1–9.
 234. Borish L, Schmaling K, DiClementi JD, Streib J, Jones JF. Chronic fatigue syndrome: identification of distinct subgroups on the basis of allergy and psychological variables. *J Allergy Clin Immunol.* 1998;102(2):222–230.
 235. Griffith JP, Zarrouf FA. A systematic review of chronic fatigue syndrome: don't assume it's depression. *Prim Care Companion J Clin Psychiatry.* 2008;10(2):120–128.
 236. Zelechowska P, Przejinska-Blaszczak E, Agier J, Kozłowska E. Different effectiveness of fungal pathogen-associated molecular patterns (PAMPs) in activating rat peritoneal mast cells. *Immunol Lett.* 2022;248:7–15.
 237. Tan J, Anderson DE, Rathore APS, et al. Signatures of mast cell activation are associated with severe COVID-19. *medRxiv: the preprint server for health sciences.* 2021. <https://doi.org/10.1101/2021.05.31.21255594>.

238. Wu ML, Liu FL, Sun J, Li X, He XY, Zheng HY, et al. SARS-CoV-2-triggered mast cell rapid degranulation induces alveolar epithelial inflammation and lung injury. *Signal Transduct Target Ther*. 2021;6(1):428.
239. Helms J, Kremer S, Merdji H, Clere-Jehl R, Schenck M, Kummerlen C, et al. Neurologic features in severe SARS-CoV-2 infection. *N Engl J Med*. 2020;382(23):2268–2270.
240. Nazari S, Azari Jafari A, Mirmoeeni S, Sadeghian S, Heidari ME, Sadeghian S, et al. Central nervous system manifestations in COVID-19 patients: a systematic review and meta-analysis. *Brain Behav*. 2021;11(5):e02025.
241. Wechsler JB, Butuci M, Wong A, Kamboj AP, Youngblood BA. Mast cell activation is associated with post-acute COVID-19 syndrome. *Allergy*. 2022;77(4):1288–1291.
242. Krysko O, Bourne JH, Kondakova E, Galova EA, Whitworth K, Newby ML, et al. Severity of SARS-CoV-2 infection is associated with high numbers of alveolar mast cells and their degranulation. *Front Immunol*. 2022;13: 968981.
243. Motta Junior JDS, Miggiolaro A, Nagashima S, de Paula CBV, Baena CP, Scharfstein J, et al. Mast cells in alveolar septa of COVID-19 patients: a pathogenic pathway that may link interstitial edema to immunothrombosis. *Front Immunol*. 2020;11: 574862.
244. Al-Rawi ZS, Gorial FI, Menhad A. Joint hypermobility and joint hypermobility syndrome in Iraqi patients with asthma. *Int J Mod Biol Med*. 2012;2(1):39–45.
245. Kohno R, Cannom DS, Olshansky B, Xi SC, Krishnappa D, Adkisson WO, et al. Mast cell activation disorder and postural orthostatic tachycardia syndrome: a clinical association. *J Am Heart Assoc*. 2021;10(17): e021002.
246. Monaco A, Choi D, Uzun S, Maitland A, Riley B. Association of mast-cell-related conditions with hypermobile syndromes: a review of the literature. *Immunol Res*. 2022;70(4):419–431.
247. Szaletowski RJ, Davis BP. Ehlers-Danlos Syndrome is associated with Idiopathic Urticaria-a retrospective study [abstract]. *J Allergy Clin Immunol*. 2019;143. <https://doi.org/10.1016/j.jaci.2018.12.204>.
248. Kohn A, Chang C. The relationship between hypermobile Ehlers-Danlos syndrome (hEDS), postural orthostatic tachycardia syndrome (POTS), and mast cell activation syndrome (MCAS). *Clin Rev Allergy Immunol*. 2020;58(3):273–297.
249. Braconnier ML, Siper PM. Neuropsychological assessment in autism spectrum disorder. *Curr Psychiatry Rep*. 2021;23(10):63.
250. Kirby AV, Bilder DA, Wiggins LD, Hughes MM, Davis J, Hall-Lande JA, et al. Sensory features in autism: findings from a large population-based surveillance system. *Autism Res*. 2022;15(4):751–760.
251. Shenouda J, Barrett E, Davidow AL, Halperin W, Silenzio VMB, Zahorodny W. Prevalence of autism spectrum disorder in a large, diverse metropolitan area: variation by sociodemographic factors. *Autism Res*. 2022;15(1):146–155.
252. Etyemez S, Esler A, Kini A, Tsai PC, DiRienzo M, Maenner M, et al. The role of intellectual disability with autism spectrum disorder and the documented cooccurring conditions: a population-based study. *Autism Res*. 2022;15(12):2399–2408.
253. Theoharides TC, Stewart JM, Panagiotidou S, Melamed I. Mast cells, brain inflammation and autism. *Eur J Pharmacol*. 2016;778:96–102.
254. Theoharides TC, Kavaloti M, Tsiloni I. Mast cells, stress, fear and autism spectrum disorder. *Int J Mol Sci*. 2019;20(15):3611.
255. Theoharides TC. Autism spectrum disorders and mastocytosis. *Int J Immunopathol Pharmacol*. 2009;22(4):859–865.
256. Ravn NH, Halling AS, Berkowitz AG, Rinnov MR, Silverberg JJ, Egeberg A, et al. How does parental history of atopic disease predict the risk of atopic dermatitis in a child? A systematic review and meta-analysis. *J Allergy Clin Immunol*. 2020;145(4):1182–1193.
257. Estes ML, McAllister AK. Maternal immune activation: implications for neuropsychiatric disorders. *Science*. 2016;353(6301):772–777.
258. Zerbo O, Leong A, Barcellos L, Bernal P, Fireman B, Croen LA. Immune mediated conditions in autism spectrum disorders. *Brain Behav Immun*. 2015;46:232–236.
259. Croen LA, Grether JK, Yoshida CK, Odouli R, Van de Water J. Maternal autoimmune diseases, asthma and allergies, and childhood autism spectrum disorders: a case-control study. *Arch Pediatr Adolesc Med*. 2005;159(2):151–157.
260. Comi AM, Zimmerman AW, Frye VH, Law PA, Peeden JN. Familial clustering of autoimmune disorders and evaluation of medical risk factors in autism. *J Child Neurol*. 1999;14(6):388–394.
261. Patel S, Cooper MN, Jones H, Whitehouse AJO, Dale RC, Guastella AJ. Maternal immune-related conditions during pregnancy may be a risk factor for neuropsychiatric problems in offspring throughout childhood and adolescence. *Psychol Med*. 2021;51(16):2904–2914.
262. Msallam R, Balla J, Rathore APS, Kared H, Malleret B, Saron WAA, et al. Fetal mast cells mediate postnatal allergic responses dependent on maternal IgE. *Science*. 2020;370(6519):941–950.
263. Theoharides TC, Petra AI, Taracanov A, Panagiotidou S, Conti P. Targeting IL-33 in autoimmunity and inflammation. *J Pharmacol Exp Ther*. 2015;354(1):24–31.
264. Angelidou A, Asadi S, Alysandratos KD, Karagkouni A, Kourembanas S, Theoharides TC. Perinatal stress, brain inflammation and risk of autism-review and proposal. *BMC Pediatr*. 2012;12:89.
265. Mottahedin A, Ardalan M, Chumak T, Riebe I, Ek J, Mallard C. Effect of neuroinflammation on synaptic organization and function in the developing brain: implications for neurodevelopmental and neurodegenerative disorders. *Front Cell Neurosci*. 2017;11:190.
266. Hagberg H, Gressens P, Mallard C. Inflammation during fetal and neonatal life: implications for neurologic and neuropsychiatric disease in children and adults. *Ann Neurol*. 2012;71(4):444–457.
267. Kelly SB, Dean JM, Zahra VA, Dudink I, Thiel A, Polglase GR, et al. Progressive inflammation reduces high-frequency EEG activity and cortical dendritic arborisation in late gestation fetal sheep. *J Neuroinflammation*. 2023;20(1):124.
268. Lenz KM, Pickett LA, Wright CL, Davis KT, Joshi A, McCarthy MM. Mast cells in the developing brain determine adult sexual behavior. *J Neurosci*. 2018;38(37):8044–8059.
269. Chang HY, Seo JH, Kim HY, Kwon JW, Kim BJ, Kim HB, et al. Allergic diseases in preschoolers are associated with psychological and behavioural problems. *Allergy Asthma Immunol Res*. 2013;5(5):315–321.
270. Johnson CP, Myers SM. American Academy of Pediatrics Council on Children With Disabilities. Identification and evaluation of children with autism spectrum disorders. *Pediatrics*. 2007;120(5):1183–1215.
271. Lai MC, Lombardo MV, Baron-Cohen S. Autism. *Lancet*. 2014;383(9920):896–910.
272. Xu G, Strathearn L, Liu B, Bao W. Prevalence of autism spectrum disorder among US children and adolescents, 2014–2016. *JAMA*. 2018;319(1):81–82.
273. McPartland J, Volkmar FR. Autism and related disorders. *Handb Clin Neurol*. 2012;106:407–418.
274. Liao TC, Lien YT, Wang S, Huang SL, Chen CY. Comorbidity of atopic disorders with autism spectrum disorder and attention deficit/hyperactivity disorder. *J Pediatr*. 2016;171:248–255.
275. Theoharides TC. Is a subtype of autism an allergy of the brain? *Clin Ther*. 2013;35(5):584–591.
276. Qu X, Lee LC, Ladd-Acosta C, Hong X, Ji Y, Kalb LG, et al. Association between atopic diseases and neurodevelopmental disabilities in a longitudinal birth cohort. *Autism Res*. 2022;15(4):740–750.
277. Jameson C, Boulton KA, Silove N, Guastella AJ. Eczema and related atopic diseases are associated with increased symptom severity in children with autism spectrum disorder. *Transl Psychiatry*. 2022;12(1):415.
278. Xu G, Snetelaar LG, Jing J, Liu B, Strathearn L, Bao W. Association of food allergy and other allergic conditions with autism spectrum disorder in children. *JAMA Netw Open*. 2018;1(2): e180279.
279. Tan Y, Thomas S, Lee BK. Parent-reported prevalence of food allergies in children with autism spectrum disorder: national health interview survey, 2011–2015. *Autism Res*. 2019;12(5):802–805.
280. Peretti S, Mariano M, Mazzocchi C, Mazza M, Pino MC, Verrotti Di Pianella A, et al. Diet: the keystone of autism spectrum disorder? *Nutr Neurosci*. 2019;22(12):825–839.
281. Mostafa GA, Al-Ayadhi LY. The possible relationship between allergic manifestations and elevated serum levels of brain specific auto-antibodies in autistic children. *J Neuroimmunol*. 2013;261(1–2):77–81.
282. Cortes Rivera M, Mastronardi C, Silva-Aldana CT, Arcos-Burgos M, Lidbury BA. Myalgic encephalomyelitis/chronic fatigue syndrome: a comprehensive review. *Diagnostics (Basel)*. 2019;9(3):91.
283. Yancey JR, Thomas SM. Chronic fatigue syndrome: diagnosis and treatment. *Am Fam Phys*. 2012;86(8):741–746.
284. Nguyen T, Johnston S, Chacko A, Gibson D, Cepon J, Smith P, et al. Novel characterisation of mast cell phenotypes from peripheral blood mononuclear cells in chronic fatigue syndrome/myalgic encephalomyelitis patients. *Asian Pac J Allergy Immunol*. 2017;35(2):75–81.
285. Bateman L, Bested AC, Bonilla HF, Chheda BV, Chu L, Curtin JM, et al. Myalgic encephalomyelitis/chronic fatigue syndrome: essentials of diagnosis and management. *Mayo Clin Proc*. 2021;96(11):2861–2878.
286. Deumer US, Varesi A, Floris V, Savioli G, Mantovani E, López-Carrasco P, et al. Myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS): an overview. *J Clin Med*. 2021;10(20):4786.
287. Ceban F, Ling S, Lui LMW, Lee Y, Gill H, Teopiz KM, et al. Fatigue and cognitive impairment in post-COVID-19 Syndrome: a systematic review and meta-analysis. *Brain Behav Immun*. 2022;101:93–135.
288. van't Leven M, Zielhuis GA, van der Meer JW, Verbeek AL, Bleijenberg G. Fatigue and chronic fatigue syndrome-like complaints in the general population. *Eur J Public Health*. 2010;20(3):251–257.
289. Natelson BH. Myalgic encephalomyelitis/chronic fatigue syndrome and fibromyalgia: definitions, similarities, and differences. *Clin Ther*. 2019;41(4):612–618.
290. Jason LA, Corradi K, Torres-Harding S, Taylor RR, King C. Chronic fatigue syndrome: the need for subtypes. *Neuropsychol Rev*. 2005;15(1):29–58.
291. Anderson VR, Jason LA, Hlavaty LE, Porter N, Cudia J. A review and meta-synthesis of qualitative studies on myalgic encephalomyelitis/chronic fatigue syndrome. *Patient Educ Couns*. 2012;86(2):147–155.
292. Ciccone DS, Natelson BH. Comorbid illness in women with chronic fatigue syndrome: a test of the single syndrome hypothesis. *Psychosom Med*. 2003;65(2):268–275.
293. Wu TY, Khorramshahi T, Taylor LA, Bansal NS, Rodriguez B, Rey IR. Prevalence of Aspergillus-derived mycotoxins (ochratoxin, aflatoxin, and gliotoxin) and their distribution in the urinalysis of ME/CFS patients. *Int J Environ Res Public Health*. 2022;19(4):2052.
294. Theoharides TC, Weinkauff C, Conti P. Brain cytokines and neuropsychiatric disorders. *J Clin Psychopharmacol*. 2004;24(6):577–581.
295. Sotzny F, Blanco J, Capelli E, Castro-Marrero J, Steiner S, Murovska M, et al. Myalgic Encephalomyelitis/Chronic Fatigue Syndrome - Evidence for an autoimmune disease. *Autoimmun Rev*. 2018;17(6):601–609.
296. Dietert RR, Dietert JM. Possible role for early-life immune insult including developmental immunotoxicity in chronic fatigue syndrome (CFS) or myalgic encephalomyelitis (ME). *Toxicology*. 2008;247(1):61–72.
297. Bower JE. Fatigue, brain, behavior, and immunity: summary of the 2012 Named Series on Fatigue. *Brain Behav Immun*. 2012;26(8):1220–1223.
298. Nakatomi Y, Mizuno K, Ishii A, Wada Y, Tanaka M, Tazawa S, et al. Neuroinflammation in patients with chronic fatigue syndrome/myalgic encephalomyelitis: an ¹¹C-(R)-PK11195 PET Study. *J Nucl Med*. 2014;55(6):945–950.

299. Keykavousi K, Nourbakhsh F, Abdollahpour N, Fazeli F, Sedaghat A, Soheili V, et al. A review of routine laboratory biomarkers for the detection of severe COVID-19 disease. *Int J Anal Chem*. 2022;2022: 9006487.
300. Cron RQ, Goyal G, Chatham WW. Cytokine storm syndrome. *Annu Rev Med*. 2023;74:321–337.
301. Dutta D, Liu J, Xiong H. NLRP3 inflammasome activation and SARS-CoV-2-mediated hyperinflammation, cytokine storm and neurological syndromes. *Int J Physiol Pathophysiol Pharmacol*. 2022;14(3):138–160.
302. Rasool G, Riaz M, Abbas M, Fatima H, Qamar MM, Zafar F, et al. COVID-19: clinical laboratory diagnosis and monitoring of novel coronavirus infected patients using molecular, serological and biochemical markers: a review. *Int J Immunopathol Pharmacol*. 2022;36: 3946320221115316.
303. Sukocheva OA, Maksoud R, Beeraka NM, Madhunapantula SV, Sinelnikov M, Nikolenko VN, et al. Analysis of post COVID-19 condition and its overlap with myalgic encephalomyelitis/chronic fatigue syndrome. *J Adv Res*. 2022;40:179–196.
304. Fletcher MA, Zeng XR, Barnes Z, Levis S, Klimas NG. Plasma cytokines in women with chronic fatigue syndrome. *J Transl Med*. 2009;7:96.
305. Maes M, Twisk FN, Ringel K. Inflammatory and cell-mediated immune biomarkers in myalgic encephalomyelitis/chronic fatigue syndrome and depression: inflammatory markers are higher in myalgic encephalomyelitis/chronic fatigue syndrome than in depression. *Psychother Psychosom*. 2012;81(5):286–295.
306. Nakamura T, Schwander S, Donnelly R, Cook DB, Ortega F, Togo F, et al. Exercise and sleep deprivation do not change cytokine expression levels in patients with chronic fatigue syndrome. *Clin Vaccin Immunol*. 2013;20(11):1736–1742.
307. Milrad SF, Hall DL, Jutagir DR, Lattie EG, Czaja SJ, Perdomo DM, et al. Depression, evening salivary cortisol and inflammation in chronic fatigue syndrome: a psychoneuroendocrinological structural regression model. *Int J Psychophysiol*. 2018;131:124–130.
308. Giloteaux L, O'Neal A, Castro-Marrero J, Levine SM, Hanson MR. Cytokine profiling of extracellular vesicles isolated from plasma in myalgic encephalomyelitis/chronic fatigue syndrome: a pilot study. *J Transl Med*. 2020;18(1):387.
309. Klimas NG, Broderick G, Fletcher MA. Biomarkers for chronic fatigue. *Brain Behav Immun*. 2012;26(8):1202–1210.
310. Choutka J, Jansari V, Hornig M, Iwasaki A. Author Correction: unexplained post-acute infection syndromes. *Nat Med*. 2022;28(8):1723.
311. Long COVID: let patients help define long-lasting COVID symptoms. *Nature*. 2020;586(7828):170.
312. White P. Long COVID: don't consign ME/CFS to history. *Nature*. 2020;587(7833):197.
313. Kempuraj D, Selvakumar GP, Thangavel R, Ahmed ME, Zaheer S, Kumar KK, et al. Glia maturation factor and mast cell-dependent expression of inflammatory mediators and proteinase activated Receptor-2 in neuroinflammation. *J Alzheimers Dis*. 2018;66(3):1117–1129.
314. Wirth KJ, Scheibenbogen C, Paul F. An attempt to explain the neurological symptoms of myalgic encephalomyelitis/chronic fatigue syndrome. *J Transl Med*. 2021;19(1):471.
315. Haffke M, Freitag H, Rudolf G, Seifert M, Doehner W, Scherbakov N, et al. Endothelial dysfunction and altered endothelial biomarkers in patients with post-COVID-19 syndrome and chronic fatigue syndrome (ME/CFS). *J Transl Med*. 2022;20(1):138.
316. Campen C, Rowe PC, Visser FC. Orthostatic symptoms and reductions in cerebral blood flow in long-haul COVID-19 patients: similarities with myalgic encephalomyelitis/chronic fatigue syndrome. *Medicina (Kaunas)*. 2021;58(1):28.
317. Vernon SD, Funk S, Bateman L, Stoddard GJ, Hammer S, Sullivan K, et al. Orthostatic challenge causes distinctive symptomatic, hemodynamic and cognitive responses in Long COVID and myalgic encephalomyelitis/chronic fatigue syndrome. *Front Med (Lausanne)*. 2022;9: 917019.
318. Patel MA, Knauer MJ, Nicholson M, Daley M, Van Nynatten LR, Martin C, et al. Elevated vascular transformation blood biomarkers in Long-COVID indicate angiogenesis as a key pathophysiological mechanism. *Mol Med*. 2022;28(1):122.
319. Gravelina S, Vilmane A, Svirskis S, Rasa-Dzelzkaleja S, Nora-Krukke Z, Vecvagare K, et al. Biomarkers in the diagnostic algorithm of myalgic encephalomyelitis/chronic fatigue syndrome. *Front Immunol*. 2022;13: 928945.
320. Wang Z, Waldman MF, Basavanahally TJ, Jacobs AR, Lopez G, Perichon RY, et al. Autoimmune gene expression profiling of fingerstick whole blood in Chronic Fatigue Syndrome. *J Transl Med*. 2022;20(1):486.
321. Vikse J, Omdal R. Fatigue in mastocytosis: a case series. *Clin Ther*. 2019;41(4):625–632.
322. Georgin-Lavialle S, Gaillard R, Moura D, Hermine O. Mastocytosis in adulthood and neuropsychiatric disorders. *Transl Res*. 2016;174:77–85.e1.
323. Yang TY, Kuo HT, Chen HJ, Chen CS, Lin WM, Tsai SY, et al. Increased risk of chronic fatigue syndrome following atopy: a population-based study. *Medicine (Baltimore)*. 2015;94(29):e1211.
324. Viner R, Hotopf M. Childhood predictors of self reported chronic fatigue syndrome/myalgic encephalomyelitis in adults: national birth cohort study. *BMJ*. 2004;329(7472):941.
325. Straus SE, Dale JK, Wright R, Metcalfe DD. Allergy and the chronic fatigue syndrome. *J Allergy Clin Immunol*. 1988;81(5 Pt 1):791–795.
326. Tsai SY, Chen HJ, Chen C, Lio CF, Kuo CF, Leong KH, et al. Increased risk of chronic fatigue syndrome following psoriasis: a nationwide population-based cohort study. *J Transl Med*. 2019;17(1):154.
327. Tsilioni I, Natelson B, Theoharides TC. Exosome-associated mitochondrial DNA from patients with myalgic encephalomyelitis/chronic fatigue syndrome stimulates human microglia to release IL-1 β . *Eur J Neurosci*. 2022;56(10):5784–5794.
328. Lin MM, Liu N, Qin ZH, Wang Y. Mitochondrial-derived damage-associated molecular patterns amplify neuroinflammation in neurodegenerative diseases. *Acta Pharmacol Sin*. 2022;43(10):2439–2447.
329. Patel AB, Theoharides TC. Methoxyluteolin inhibits neuropeptide-stimulated proinflammatory mediator release via mTOR activation from human mast cells. *J Pharmacol Exp Ther*. 2017;361(3):462–471.
330. Wallis CM, Munyaneza JE, Chen J, Novy R, Bester G, Buchman JL, et al. Candidatus *Liberibacter solanacearum* titers in and infection effects on potato tuber chemistry of promising germplasm exhibiting tolerance to zebra chip disease. *Phytopathology*. 2015;105(12):1573–1584.
331. Mannick JB, Teo G, Bernardo P, Quinn D, Russell K, Klickstein L, et al. Targeting the biology of ageing with mTOR inhibitors to improve immune function in older adults: phase 2b and phase 3 randomised trials. *Lancet Healthy Longev*. 2021;2(5):e250–e262.
332. Morawe MP, Liao F, Amberg W, van Bergeijk J, Chang R, Gulino M, et al. Pharmacological mTOR-inhibition facilitates clearance of AD-related tau aggregates in the mouse brain. *Eur J Pharmacol*. 2022;934: 175301.
333. Girodengo M, Ultanir SK, Bateman JM. Mechanistic target of rapamycin signaling in human nervous system development and disease. *Front Mol Neurosci*. 2022;15: 1005631.
334. Xu F, Na L, Li Y, Chen L. Roles of the PI3K/AKT/mTOR signalling pathways in neurodegenerative diseases and tumours. *Cell Biosci*. 2020;10(1):54.
335. Rapaka D, Bitra VR, Challa SR. Adukuwu PC. mTOR signaling as a molecular target for the alleviation of Alzheimer's disease pathogenesis. *Neurochem Int*. 2022;155: 105311.
336. Costa-Mattoli M, Monteggia LM. mTOR complexes in neurodevelopmental and neuropsychiatric disorders. *Nat Neurosci*. 2013;16(11):1537–1543.
337. Ryskalin L, Limanaqi F, Frati A, Busceti CL, Fornai F. mTOR-related brain dysfunctions in neuropsychiatric disorders. *Int J Mol Sci*. 2018;19(8):2226.
338. Jernigan CS, Goswami DB, Austin MC, Iyo AH, Chandran A, Stockmeier CA, et al. The mTOR signaling pathway in the prefrontal cortex is compromised in major depressive disorder. *Prog Neuropsychopharmacol Biol Psychiatry*. 2011;35(7):1774–1779.
339. Lipton JO, Sahin M. The neurology of mTOR. *Neuron*. 2014;84(2):275–291.
340. Movahedpour A, Vakili O, Khalifeh M, Mousavi P, Mahmoodzadeh A, Taheri-Anganeh M, et al. Mammalian target of rapamycin (mTOR) signaling pathway and traumatic brain injury: a novel insight into targeted therapy. *Cell Biochem Funct*. 2022;40(3):232–247.
341. Xu T, Liu J, Li XR, Yu Y, Luo X, Zheng X, et al. The mTOR/NF- κ B pathway mediates neuroinflammation and synaptic plasticity in diabetic encephalopathy. *Mol Neurobiol*. 2021;58(8):3848–3862.
342. He G, Jiang H, Zhang E, Wan P, Liu G, Ji Z. Evaluating the value of p70S6K and mTOR signaling pathway in monitoring exercise-induced central fatigue in rats. *Cell Mol Biol (Noisy-Le-Grand)*. 2022;68(10):79–83.
343. Brodin P. Immune determinants of COVID-19 disease presentation and severity. *Nat Med*. 2021;27(1):28–33.
344. Ye Q, Wang B, Mao J. The pathogenesis and treatment of the 'cytokine Storm'. *J Infectol*. 2020;80(6):607–613.
345. Chen G, Wu D, Guo W, Cao Y, Huang D, Wang H, et al. Clinical and immunological features of severe and moderate coronavirus disease 2019. *J Clin Invest*. 2020;130(5):2620–2629.
346. Conti P, Ronconi G, Caraffa A, Gallenga CE, Ross R, Frydas I, et al. Induction of pro-inflammatory cytokines (IL-1 and IL-6) and lung inflammation by Coronavirus-19 (COVI-19 or SARS-CoV-2): anti-inflammatory strategies. *J Biol Regul Homeost Agents*. 2020;34(2):327–331.
347. Giamarellos-Bourboulis EJ, Netea MG, Rovina N, Rovina N, Akinosoglou K, Antoniadou A, et al. Complex immune dysregulation in COVID-19 patients with severe respiratory failure. *Cell Host Microbe*. 2020;27(6):992–1000.e3.
348. Zhang C, Baumer A, Mackay IR, Linnane AW, Nagley P. Unusual pattern of mitochondrial DNA deletions in skeletal muscle of an adult human with chronic fatigue syndrome. *Hum Mol Genet*. 1995;4(4):751–754.
349. Tang Y, Liu J, Zhang D, Xu Z, Ji J, Wen C. Cytokine storm in COVID-19: the current evidence and treatment strategies. *Front Immunol*. 2020;11:1708.
350. Paces J, Strizova Z, Smrz D, Cerny J. COVID-19 and the immune system. *Physiol Res*. 2020;69(3):379–388.
351. Ragab D, Salah Eldin H, Taeimah M, Khattab R, Salem R. The COVID-19 cytokine storm; what we know so far. *Front Immunol*. 2020;11:1446.
352. Herold T, Jurinovic V, Arnreich C, Lipworth BJ, Hellmuth JC, von Bergwelt-Baildon M, et al. Elevated levels of IL-6 and CRP predict the need for mechanical ventilation in COVID-19. *J Allergy Clin Immunol*. 2020;146(1):128–136.e4.
353. Copescu A, Smibert O, Gibson A, Phillips EJ, Trubiano JA. The role of IL-6 and other mediators in the cytokine storm associated with SARS-CoV-2 infection. *J Allergy Clin Immunol*. 2020;146(3):518–534.e1.
354. Han H, Ma Q, Li C, Liu R, Zhao L, Wang W, et al. Profiling serum cytokines in COVID-19 patients reveals IL-6 and IL-10 are disease severity predictors. *Emerg Microbes Infect*. 2020;9(1):1123–1130.
355. Mazzoni A, Salvati L, Maggi L, Capone M, Vanni A, Spinicci M, et al. Impaired immune cell cytotoxicity in severe COVID-19 is IL-6 dependent. *J Clin Invest*. 2020;130(9):4694–4703.
356. Liu F, Li L, Xu M, Wu J, Luo D, Zhu Y, et al. Prognostic value of interleukin-6, C-reactive protein, and procalcitonin in patients with COVID-19. *J Clin Virol*. 2020;127: 104370.
357. Canna SW, Cron RQ. Highways to hell: mechanism-based management of cytokine storm syndromes. *J Allergy Clin Immunol*. 2020;146(5):949–959.

358. Scozzi D, Cano M, Ma L, Zhou D, Zhu JH, O'Halloran JA, et al. Circulating mitochondrial DNA is an early indicator of severe illness and mortality from COVID-19. *JCI Insight*. 2021;6(4): e143299.
359. Theoharides TC. COVID-19, pulmonary mast cells, cytokine storms, and beneficial actions of luteolin. *BioFactors*. 2020;46(3):306–308.
360. Theoharides TC. Potential association of mast cells with coronavirus disease 2019. *Ann Allergy Asthma Immunol*. 2021;126(3):217–218.
361. Theoharides TC, Cholevas C, Polyzoidis K, Politis A. Long-COVID syndrome-associated brain fog and chemofog: luteolin to the rescue. *BioFactors*. 2021;47(2):232–241.
362. Liu S, Suzuki Y, Takemasa E, Watanabe R, Mogi M. Mast cells promote viral entry of SARS-CoV-2 via formation of chymase/spike protein complex. *Eur J Pharmacol*. 2022;930: 175169.
363. Arun S, Storan A, Myers B. Mast cell activation syndrome and the link with Long COVID. *Br J Hosp Med (Lond)*. 2022;83(7):1–10.
364. Murdaca G, Di Gioacchino M, Greco M, Borro M, Paladin F, Petrarca C, et al. Basophils and mast cells in COVID-19 pathogenesis. *Cells*. 2021;10(10):2754.
365. Afrin LB, Weinstock LB, Molderings GJ. Covid-19 hyperinflammation and post-Covid-19 illness may be rooted in mast cell activation syndrome. *Int J Infect Dis*. 2020;100:327–332.
366. Weinstock LB, Brook JB, Walters AS, Goris A, Afrin LB, Molderings GJ. Mast cell activation symptoms are prevalent in Long-COVID. *Int J Infect Dis*. 2021;112:217–226.
367. Ozdemir O, Goksu Erol AY, Dikici U. Mast cell's role in cytokine release syndrome and related manifestations of COVID-19 disease. *Curr Pharm Des*. 2022;28(40):3261–3268.
368. Gebremeskel S, Schanin J, Coyle KM, Butuci M, Luu T, Brock EC, et al. Mast cell and eosinophil activation are associated with COVID-19 and TLR-mediated viral inflammation: implications for an anti-Siglec-8 antibody. *Front Immunol*. 2021;12: 650331.
369. Budnevsky AV, Avdeev SN, Kosanovic D, Shishkina VV, Filin AA, Esaulenko DI, et al. Role of mast cells in the pathogenesis of severe lung damage in COVID-19 patients. *Respir Res*. 2022;23(1):371.
370. Schaller T, Markl B, Claus R, Sholl L, Hornick JL, Giannetti MP, et al. Mast cells in lung damage of COVID-19 autopsies: a descriptive study. *Allergy*. 2022;77(7):2237–2239.
371. Fotuhi M, Mian A, Meysami S, Raji CA. Neurobiology of COVID-19. *J Alzheimers Dis*. 2020;76(1):3–19.
372. Najjar S, Najjar A, Chong DJ, Pramanik BK, Kirsch C, Kuzniecky RI, et al. Central nervous system complications associated with SARS-CoV-2 infection: integrative concepts of pathophysiology and case reports. *J Neuroinflammation*. 2020;17(1):231.
373. Singh AK, Bhushan B, Maurya A, Mishra G, Singh SK, Awasthi R. Novel coronavirus disease 2019 (COVID-19) and neurodegenerative disorders. *Dermatol Ther*. 2020;33(4):e13591.
374. Natale NR, Lukens JR, Petri Jr WA. The nervous system during COVID-19: caught in the crossfire. *Immunol Rev*. 2022;311(1):90–111.
375. Latorre D. Autoimmunity and SARS-CoV-2 infection: unraveling the link in neurological disorders. *Eur J Immunol*. 2022;52(10):1561–1571.
376. Taquet M, Luciano S, Geddes JR, Harrison PJ. Bidirectional associations between COVID-19 and psychiatric disorder: retrospective cohort studies of 62 354 COVID-19 cases in the USA. *Lancet Psychiatry*. 2021;8(2):130–140.
377. Ongur D, Perlis R, Goff D. Psychiatry and COVID-19. *JAMA*. 2020;324(12):1149–1150.
378. Vindegaard N, Benros ME. COVID-19 pandemic and mental health consequences: systematic review of the current evidence. *Brain Behav Immun*. 2020;89:531–542.
379. Pfefferbaum B, North CS. Mental health and the Covid-19 pandemic. *N Engl J Med*. 2020;383(6):510–512.
380. Xiang YT, Yang Y, Li W, Zhang L, Zhang Q, Cheung T, et al. Timely mental health care for the 2019 novel coronavirus outbreak is urgently needed. *Lancet Psychiatry*. 2020;7(3):228–229.
381. Gordon JA, Borja SE. The COVID-19 pandemic: setting the mental health research agenda. *Biol Psychiatry*. 2020;88(2):130–131.
382. Ariza M, Cano N, Segura B, Adan A, Bargalló N, Caldú X, et al. Neuropsychological impairment in post-COVID condition individuals with and without cognitive complaints. *Front Aging Neurosci*. 2022;14: 1029842.
383. Huang C, Huang L, Wang Y, Li X, Ren L, Gu X, et al. 6-month consequences of COVID-19 in patients discharged from hospital: a cohort study. *Lancet*. 2021;397(10270):220–232.
384. Higgins V, Sohaei D, Diamandis EP, Prassas I. COVID-19: from an acute to chronic disease? Potential long-term health consequences. *Crit Rev Clin Lab Sci*. 2021;58(5):297–310.
385. Moreno-Perez O, Merino E, Leon-Ramirez JM, Andres M, Ramos JM, Arenas-Jiménez J, et al. Post-acute COVID-19 syndrome. Incidence and risk factors: a Mediterranean cohort study. *J Infectol*. 2021;82(3):378–383.
386. Townsend L, Dyer AH, Jones K, Dunne J, Mooney A, Gaffney F, et al. Persistent fatigue following SARS-CoV-2 infection is common and independent of severity of initial infection. *PLoS One*. 2020;15(11): e0240784.
387. Zingaropoli MA, Iannetta M, Piermatteo L, Pasculli P, Latronico T, Mazzuti L, et al. Neuro-axonal damage and alteration of blood-brain barrier integrity in COVID-19 patients. *Cells*. 2022;11(16):2480.
388. Etter MM, Martins TA, Kulsvehagen L, Pössnecker E, Duchemin W, Hogan S, et al. Severe Neuro-COVID is associated with peripheral immune signatures, autoimmunity and neurodegeneration: a prospective cross-sectional study. *Nat Commun*. 2022;13(1):6777.
389. Bonetto V, Pasetto L, Lisi I, Carbonara M, Zangari R, Ferrari E, et al. Markers of blood-brain barrier disruption increase early and persistently in COVID-19 patients with neurological manifestations. *Front Immunol*. 2022;13: 1070379.
390. Lee MH, Perl DP, Nair G, Li W, Maric D, Murray H, et al. Microvascular injury in the brains of patients with Covid-19. *N Engl J Med*. 2021;384(5):481–483.
391. Needham EJ, Ren AL, Digby RJ, Norton EJ, Ebrahimi S, Outtrim JG, et al. Brain injury in COVID-19 is associated with dysregulated innate and adaptive immune responses. *Brain*. 2022;145(11):4097–4107.
392. Files JK, Sarkar S, Fram TR, Boppana S, Sterrett S, Qin K, et al. Duration of post-COVID-19 symptoms is associated with sustained SARS-CoV-2-specific immune responses. *JCI Insight*. 2021;6(15): e151544.
393. Herrera JE, Niehaus WN, Whiteson J, Azola A, Baratta JM, Fleming TK, et al. Multidisciplinary collaborative consensus guidance statement on the assessment and treatment of fatigue in postacute sequelae of SARS-CoV-2 infection (PASC) patients. *PM R*. 2021;13(9):1027–1043.
394. Paul BD, Lemle MD, Komaroff AL, Snyder SH. Redox imbalance links COVID-19 and myalgic encephalomyelitis/chronic fatigue syndrome. *Proc Natl Acad Sci U S A*. 2021;118(34): e2024358118.
395. Theoharides TC, Conti P. COVID-19 and multisystem inflammatory syndrome, or is it mast cell activation syndrome? *J Biol Regul Homeost Agents*. 2020;34(5):1633–1636.
396. Akin C, Valent P, Metcalfe DD. Mast cell activation syndrome: proposed diagnostic criteria. *J Allergy Clin Immunol*. 2010;126(6): 1099–104.e4.
397. Theoharides TC, Tsilioni I, Ren H. Recent advances in our understanding of mast cell activation - or should it be mast cell mediator disorders? *Expert Rev Clin Immunol*. 2019;15(6):639–656.
398. Theoharides TC, Tsilioni I. Humoral innate immunity and acute-phase proteins. *N Engl J Med*. 2023;388(18):1725.
399. Tsilioni I, Theoharides TC. Recombinant SARS-CoV-2 spike protein stimulates secretion of chymase, trypsinase, and IL-1 β from human mast cells, augmented by IL-33. *Int J Mol Sci*. 2023;24(11):9487.
400. Tsilioni I, Theoharides TC. Recombinant SARS-CoV-2 spike protein and its receptor binding domain stimulate release of different pro-inflammatory mediators via activation of distinct receptors on human microglia cells. *Mol Neurobiol*. 2023;60(11):6704–6714.
401. Proal AD, VanElzakker MB, Aleman S, et al. SARS-CoV-2 reservoir in post-acute sequelae of COVID-19 (PASC). *Nat. Immunol*. 2023;24(10):1616–1627. <https://doi.org/10.1038/s41590-023-01601-2>.
402. Swank Z, Senussi Y, Manickas-Hill Z, et al. Persistent circulating severe acute respiratory syndrome coronavirus 2 spike is associated with post-acute coronavirus disease 2019 sequelae. *Clinical Infectious Diseases: An Official Publication of the Infectious Diseases Society of America*. 2023;76(3):e487–e490. <https://doi.org/10.1093/cid/ciac722>.
403. Patterson BK, Francisco EB, Yogendra R, et al. Persistence of SARS-CoV-2 S1 Protein in CD16+ Monocytes in Post-Acute Sequelae of COVID-19 (PASC) up to 15 Months Post-Infection. *Front. Immunol*. 2022;12: 746021. <https://doi.org/10.3389/fimmu.2021.746021>.
404. Files JK, Sarkar S, Fram TR, et al. Duration of post-COVID-19 symptoms is associated with sustained SARS-CoV-2-specific immune responses. *JCI Insight*. 2021;6(15): e151544. <https://doi.org/10.1172/jci.insight.151544405>.
405. Rong Z, Mai M, KS, Puelles VG, Czogalla J, Schadler J, Vering J, Delbridge C, Steinke H, Frenzel H, Schmidt K, Caliskan OS, Ali M, Kolabas I, Ulukaya S, Horvath I, Zhao S, Krahmer N, Tahorpvc S, Yildirim O, Huber TB, Ondruschke B, Bechmann I, Ebert G, Protzer U, Bhatia HS, Hella F, Erturk A. SARS-CoV-2 Spike Protein Accumulation in the Skull-Meninges-Brain Axis: Potential Implications for Long-Term Neurological Complications in post-COVID-19. *BioRxiv*. 2023. <https://doi.org/10.1101/2023.04.04.535604>.
406. Tziastoudi M, Cholevas C, Stefanidis I, Theoharides TC. Genetics of COVID-19 and myalgic encephalomyelitis/chronic fatigue syndrome: a systematic review. *Ann Clin Transl Neurol*. 2022;9(11):1838–1857.
407. Keller RH, Lane JL, Klimas N, Reiter WM, Fletcher MA, van Riel F, et al. Association between HLA class II antigens and the chronic fatigue immune dysfunction syndrome. *Clin Infect Dis*. 1994;18(suppl 1):S154–S156.
408. James LM, Georgopoulos AP. At the Root of 3 "Long" Diseases: Persistent Antigens Inflicting Chronic Damage on the Brain and Other Organs in Gulf War Illness, Long-COVID-19, and Chronic Fatigue Syndrome. *Neurosci Insights*. 2022;17: 26331055221114817.
409. Theoharides TC. Could SARS-CoV-2 spike protein be responsible for long-COVID syndrome? *Mol Neurobiol*. 2022;59(3):1850–1861.
410. Garcia-Epelboim A, Christian KM. Modeling neuro-immune interactions using human pluripotent stem cells. *Curr Opin Neurobiol*. 2023;79: 102672.
411. Yip S, Wang N, Sugimura R. Give them vasculature and immune cells - how to fill the gap of organoids [e-pub ahead of print]. *Cells Tissues Organs*. <https://doi.org/10.1159/000529431>, accessed October 17, 2023.
412. Ostermann PN, Schaaf H. Human brain organoids to explore SARS-CoV-2-induced effects on the central nervous system. *Rev Med Virol*. 2023;33(2):e2430.
413. de Toledo MAS, Fu X, Maie T, Buhl EM, Götz K, Schmitz S, et al. KIT D816V mast cells derived from induced pluripotent stem cells recapitulate systemic mastocytosis transcriptional profile. *Int J Mol Sci*. 2023;24(6):5275.